

Radiochemistry in nuclear medicine

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Do not say “contrast media” (but *molecular imaging*)

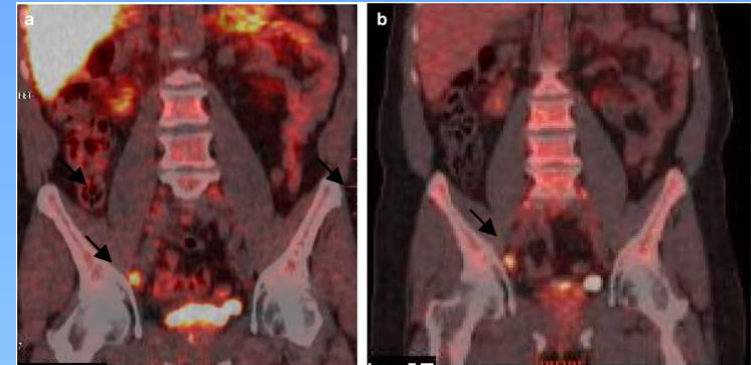
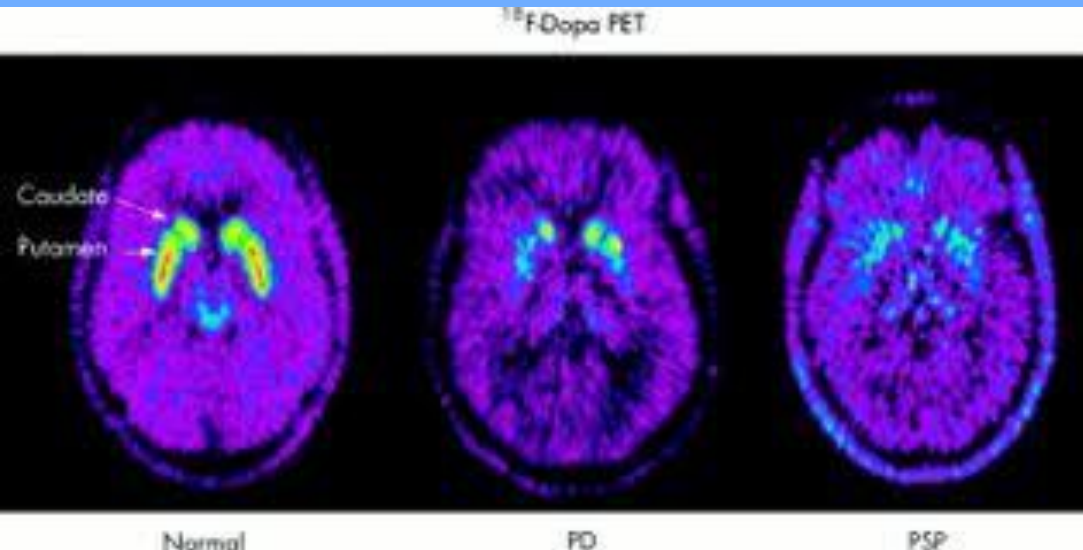
Contrast media

- Mainly anatomical
- High injected mass
 - Mild invasive
 - Patient reactions
- Non natural molecules
- Stability issues
- Easy purchase media
- MRI, CT

Radiopharmaceuticals

- Mainly functional
- Low injected mass
 - Low invasive
 - Rare patient reactions
- Some natural molecules or analogues
- Few stability issues
- Limited purchase or GMP prepared tracers
- PET, SPECT

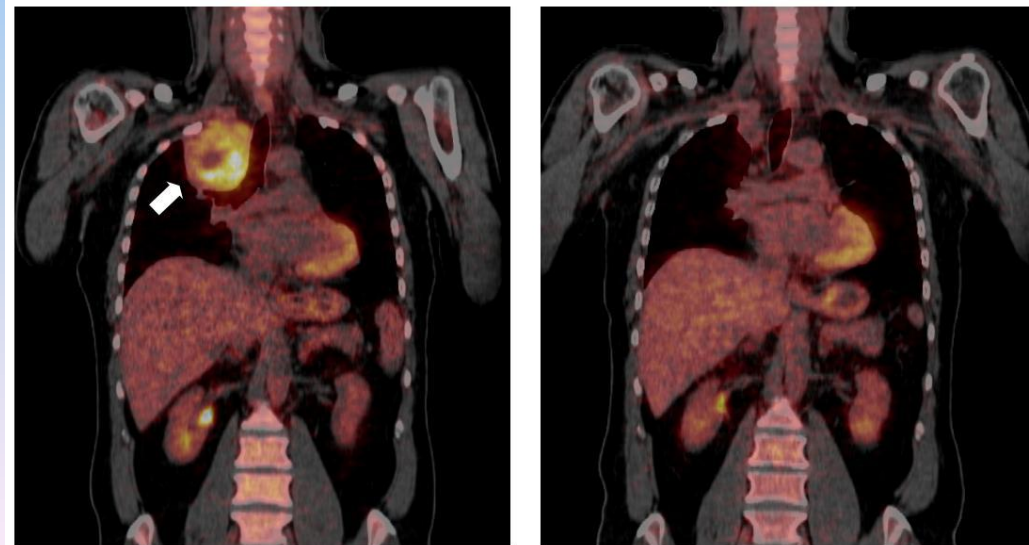
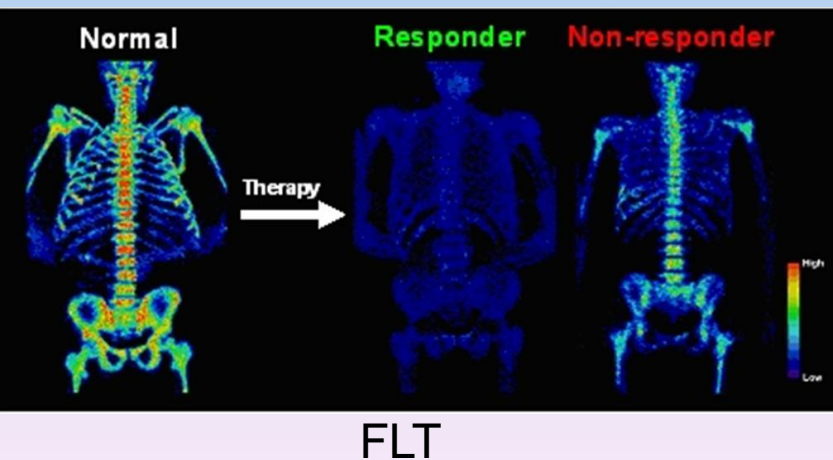
The final aim of Nuclear Medicine



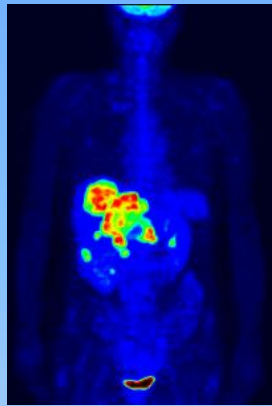
FDG

Fluorocholeline

FDG



A successful PET study



Data processing,
medical diagnosis

Physician



Patient handling,
scanning protocols

Physician
Paramedics
Physic
Engineer



Tracer radiosynthesis

Chemist
Engineer



Nuclide production

Physic
Engineer

**Big interplay among different
fields of knowledge**

Outlook

- Radionuclides in nuclear medicine
- Radiochemical reactions
- Radiopharmaceuticals

Employed radionuclides

Elements	For PET			For SPECT		
	Radionuclide	$T_{1/2}$	β^+ (MeV)	Radionuclide	$T_{1/2}$	γ (MeV)
Organic compound elements	^{11}C	20.4 months	0.959			
	^{13}N	9.96 months	1.197			
	^{15}O	2.03	1.738			
Halogens	^{18}F	109.8 months	0.635			
	^{75}Br	98 months	1.74			
	^{76}Br	16.1 h	3.98			
	^{124}I	4.2 days	2.13	^{123}I	13.2h	0.159
Metals	^{66}Ga	9.45 h	4.153	^{67}Ga	78.2h	0.093, 0.184, 0.300
	^{68}Ga	68.3 months	1.898			
	^{83}Sr	32.4	1.150			
Transition metals	^{110}In	66 months	2.250	^{111}In	67.2h	0.173, 0.247
	^{44}Sc	3.92h	1.470			
	^{47}Ti	3.09h	1.040			
	^{51}Mn	46.2 months	2.170			
	^{52}Mn	5.6 days	0.575			
	^{52}Fe	8.2h	0.800			
	^{55}Co	17. h	1.500			
	^{61}Cu	3.32h	1.220	^{67}Cu	2.6 days	0.185, 0.92
	^{62}Cu	9.76 months	2.910			
	^{64}Cu	12.8h	0.656			
	^{86}Y	14.74h	3.150			
	^{89}Zr	78.4h	0.900			
	^{94m}Tc	52 months	2.440	^{99m}Tc	6.0h	0.140

**months = minutes
Error in table!!!**

Singol Photon Emission Computed Tomography (SPECT) nuclides

^{99m}Tc ($T_{1/2}=6.02\text{h}$, $E_{\gamma}=140\text{ keV}$) is used in more than 70% of all medical applications in many pharmaceutical preparations

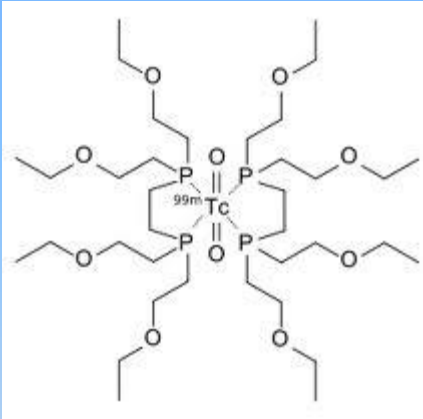
^{67}Ga ($T_{1/2}=78.3\text{h}$, $E_{\gamma}=93\text{ keV}$, 185 keV , 300 keV) is often used as tumor localizing agent (gallium citrate)

^{123}I ($T_{1/2}=13\text{h}$, $E_{\gamma}=159\text{ keV}$) can be covalently bound to several molecules and proteins. It has replaced ^{131}I ($T_{1/2}=6\text{d}$, $E_{\gamma}=364\text{ keV}$) because of the reduced radiation exposure

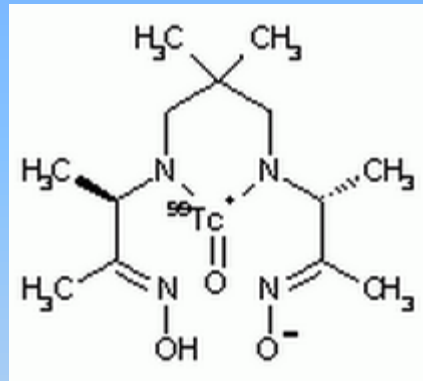
^{81m}Kr ($T_{1/2}=13\text{s}$, $E_{\gamma}=190\text{ keV}$) is a very short-lived gas used to perform lung ventilation studies, (short half-life limits its application)

Some Tc-based radiopharmaceuticals

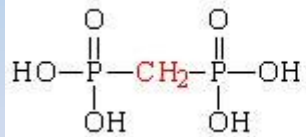
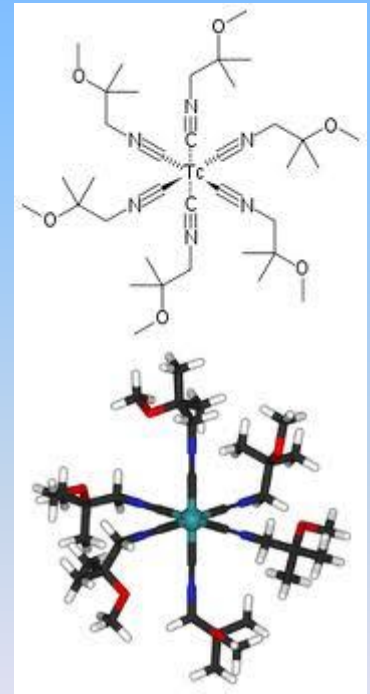
Tc-tetraphosmin (Myoview)



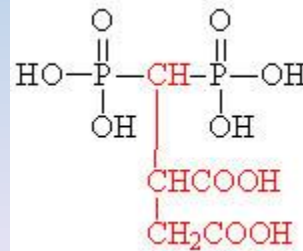
Tc-HMPAO (Ceretek)



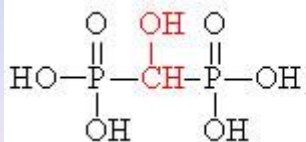
Tc-Sestamibi (Cardiolite)



MDP



DPD



HDP

Bone scan tracers

PET working principle

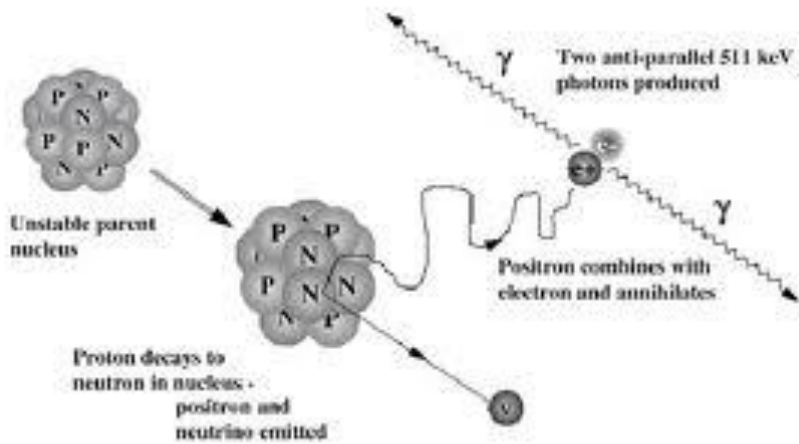
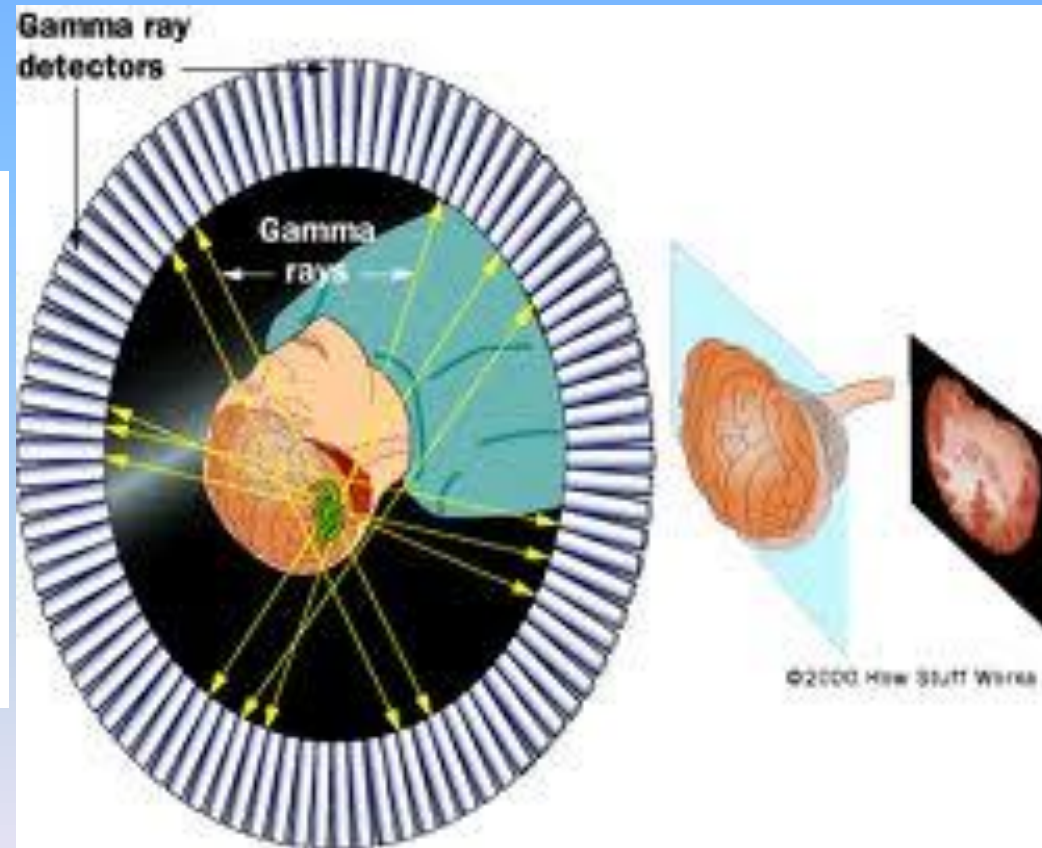


Figure 1.1. Positron emission and annihilation.



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Positron Emission Tomography (PET) nuclides

Important for PET studies are neutron deficient isotopes which decay by positron emission. Positrons annihilate with electrons emitting two $E_{\gamma}=511$ keV photons in opposite direction.

^{18}F ($T_{1/2}=110$ m, $E_{\gamma} = 511$ keV), (used in more than 80% of all PET applications)

^{13}N ($T_{1/2}=10$ m, $E_{\gamma} = 511$ keV)

^{11}C ($T_{1/2}=20.4$ m, $E_{\gamma} = 511$ keV)

^{68}Ga ($T_{1/2}=68$ m, $E_{\gamma}= 511$ keV)

^{82}Rb ($T_{1/2}=1.3$ m, $E_{\gamma} = 511$ keV)

Nuclides for radiotherapy

Radionuclide	Emission type	Half-life	E _{max} (keV)	Range in tissue	Production
¹⁸⁶ Re	β, γ (9.4%)	89.2 h	(β): 1069	Maximum (5 mm)	¹⁸⁵ Re (n, γ) ¹⁸⁶ Re
¹⁶⁶ Ho	β, γ (6.7%)	26.9 h	(β): 1853	Maximum (10.2 mm)	¹⁶⁵ Ho (n, γ) ¹⁶⁶ Ho
¹⁸⁸ Re	β, γ (15.1%)	17.0 h	(β): 2120	Maximum (11 mm)	¹⁸⁸ W/ ¹⁸⁸ Re-generator
⁸⁹ Sr	β	52.7 days	1463	Maximum (3 mm)	⁸⁸ Sr (n,γ) ⁸⁹ Sr
³² P	β	14.3 days	1710	Maximum (8.7 mm)	³² S (n,p) ³² P or ³¹ P(n, γ) ³² P
⁹⁰ Y	β	64.1 h	2280	Maximum (12 mm)	⁹⁰ Sr/ ⁹⁰ Y-generator
²²⁵ Ac	α	10 days	5830, 5792, 5790, 5732	40–80 μm	²²⁵ Ra-generaor
²¹¹ At	α	7.2 h	5870	60–80 μm	Accelerator
²¹³ Bi	α	45.7 min	5869	50–80 μm	²²⁵ Ra-generaor

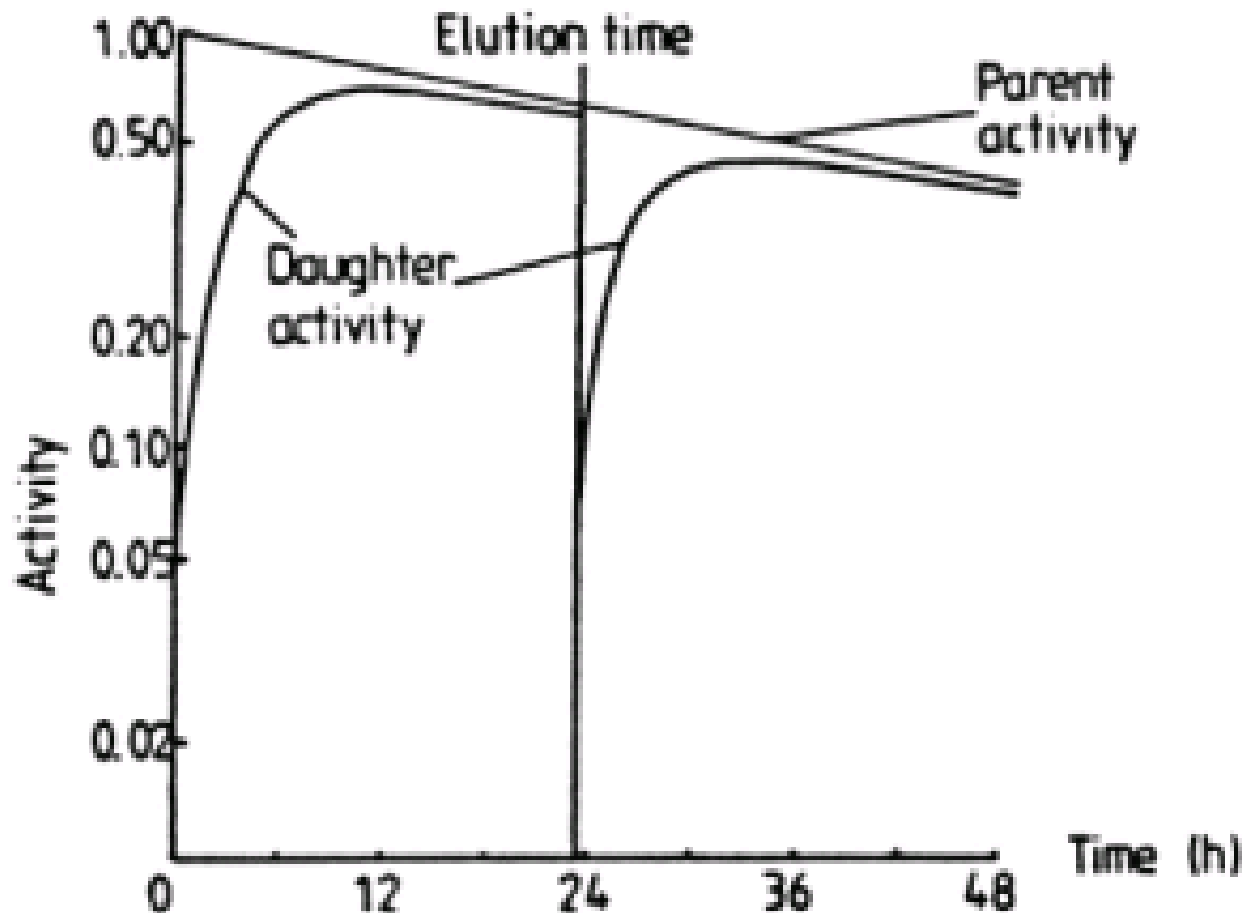
Generating radionuclides

Generators

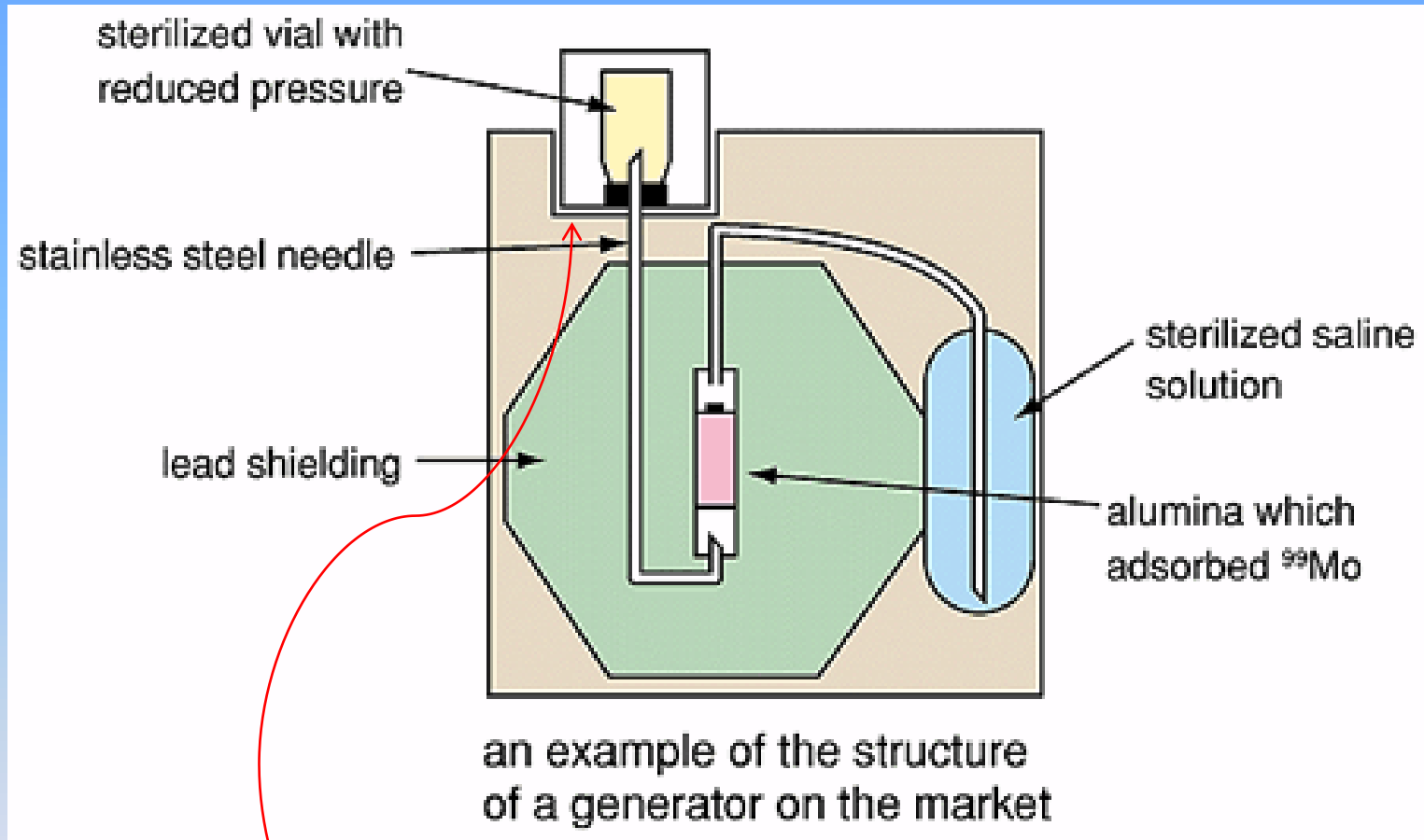
Parent P	Parent half-life	Mode of decay P→D	Daughter D	Mode of decay of D	Daughter half-life	Gamma-ray energy from daughter (keV)
⁹⁹ Mo	2.7 d	β^-	⁹⁹ Tc ^m	IT	6 h	140
⁸² Sr	25 d	EC	⁸² Rb	EC β^+	1.3 min	777 511
⁶⁸ Ge	280 d	EC	⁶⁸ Ga	EC β^+	68 min	511
⁵² Fe	8.2 h	EC β^+	⁵² Mn ^m	EC β^+ IT	21 min	511
⁸¹ Rb	4.7 h	EC	⁸¹ Kr ^m	IT	13 s	190
⁶² Zn	9.1 h	EC β^+	⁶² Cu	EC β^+	9.8 min	511
¹⁷⁸ W	21.5 d	EC	¹⁷⁸ Ta	EC	9.5 min	93

Generator elution principle

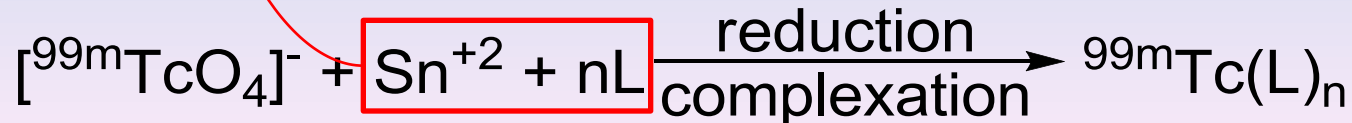
$$A_2(t) = A_1^0 \cdot [e^{-\lambda_1 \cdot t} - e^{-\lambda_2 \cdot t}]$$



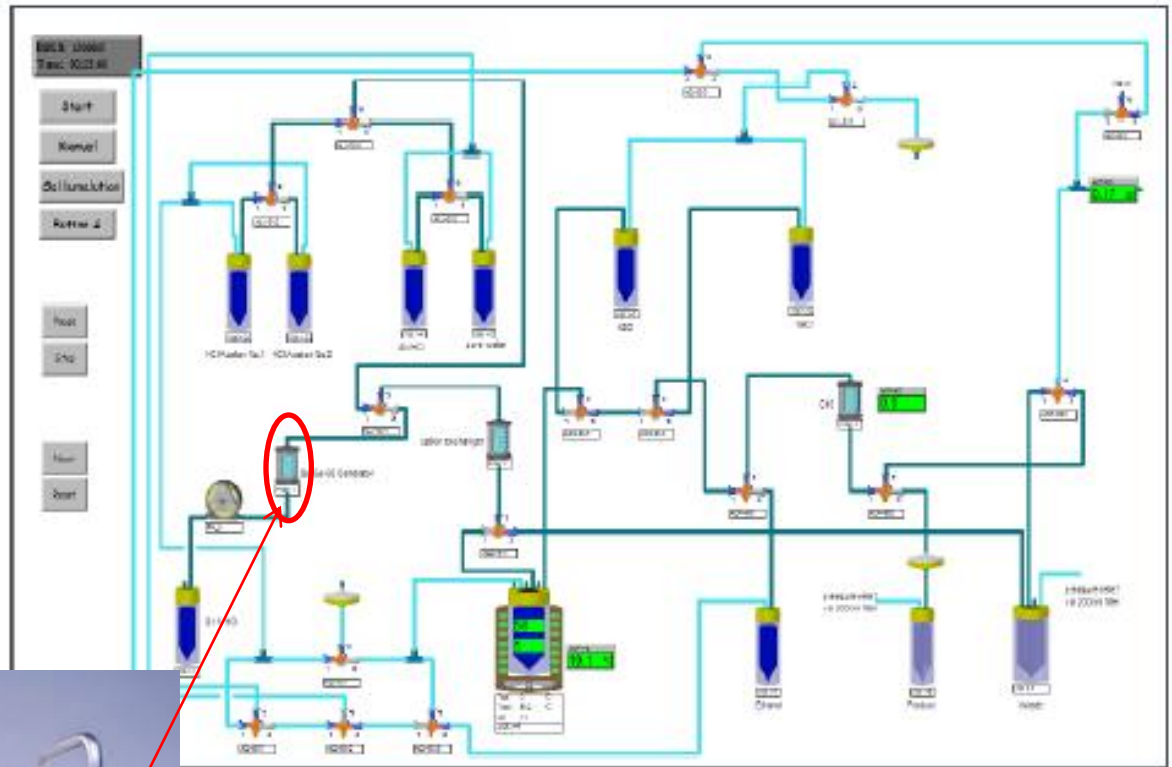
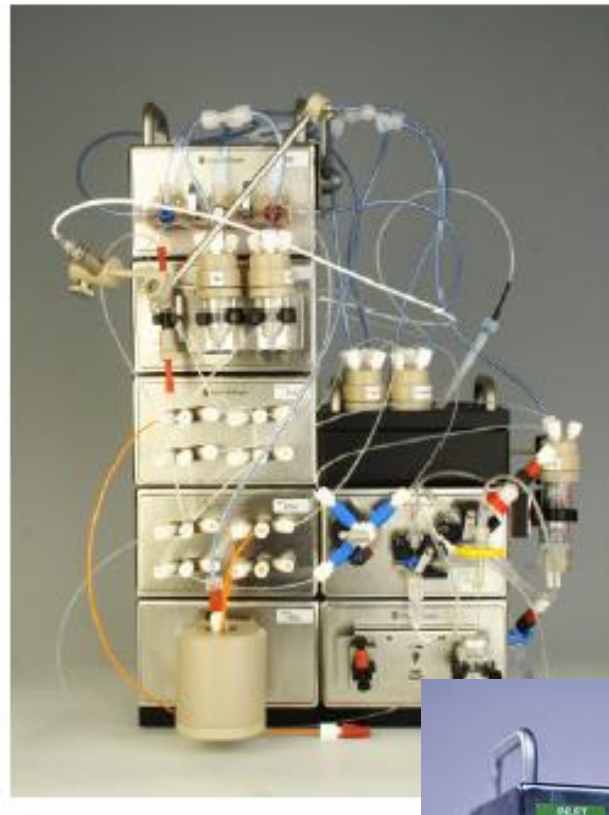
99mTc Generator Scheme



The reaction vial is ready for instantaneous preparation of tracer

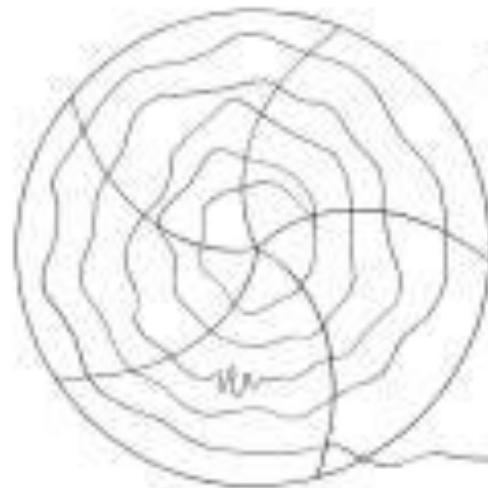


68Ga Generator and utilization



68Ga-DOTATOC synthesis apparatus scheme

Cyclotron as seen by



$$r = r_0 \left[r = \left(\frac{2qV}{m\omega} \right) \cos(2\theta - \delta_1 + \delta_2, r) + \left(\frac{2qV}{m\omega} \right) \cos(5\theta + \delta_1 - \delta_2, r) + \left(\frac{2qV}{m\omega} \right) \cos(7\theta + \delta_1 - \delta_2, r) + \dots \right] \times \left(\frac{1}{1 - \left(\frac{v}{c} \right)^2} \right)^{1/4}$$

$$\frac{d\theta}{dt} = \left[\sin(\omega t - \delta) - \sin(\omega t - \delta) \right] \frac{1}{1 - \left(\frac{v}{c} \right)^2} \times \frac{1}{\omega}$$

... the theoretical physicist

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... the student

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Biomedical cyclotron generated nuclides

Isotope	Principal gamma-ray energy (keV)	Half-life	Reaction
^{11}C	511 (β^+)	20.4 min	$^{14}\text{N}(p,\alpha)^{11}\text{C}$
^{13}N	511 (β^+)	9.96 min	$^{13}\text{C}(p,n)^{13}\text{N}$
^{15}O	511 (β^+)	2.07 min	$^{15}\text{N}(p,n)^{15}\text{O}$
^{18}F	511 (β^+)	109.7 min	$^{18}\text{O}(p,n)^{18}\text{F}$
^{67}Ga	93	78.3 h	$^{68}\text{Zn}(p,2n)^{67}\text{Ga}$
	184		
	300		
^{111}In	171	67.9 h	$^{112}\text{Cd}(p,2n)^{111}\text{In}$
	245		
^{123}I	159	13 h	$^{124}\text{Te}(p,2n)^{123}\text{I}$
			$^{127}\text{I}(p,5n)^{123}\text{Xe} \rightarrow ^{123}\text{I}$
^{201}Tl	68–80.3	73 h	$^{203}\text{Tl}(p,3n)^{201}\text{Pb} \rightarrow ^{201}\text{Tl}$

$^{94\text{m}}\text{Tc}$, ^{76}Br , ^{60}Cu , ^{64}Cu

Production example: ^{64}Cu

Cyclotron reaction
 $^{64}\text{Ni}(p,n)^{64}\text{Cu}$

6N HCl dissolution
of target material

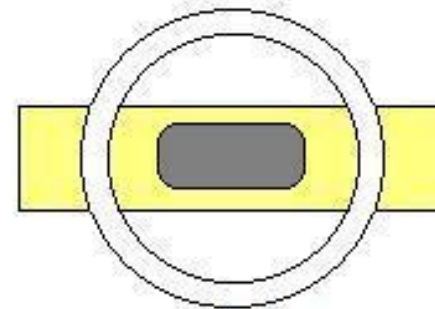
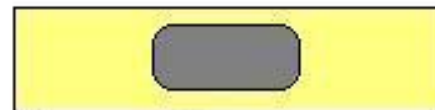
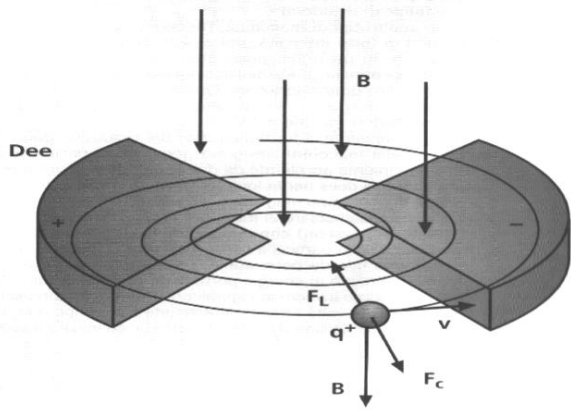
^{64}Cu separation by elution
through AG1-X8 column

$^{64}\text{CuCl}_2$ aqueous solution

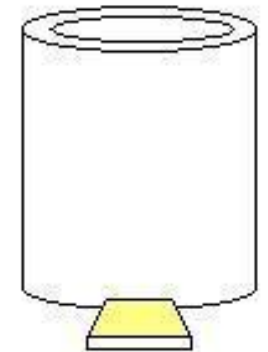
Slant target



- ^{64}Ni
- Ni nat
- Au
- Cu nat



Dissolution and electrodeposition
cylinder (up view)



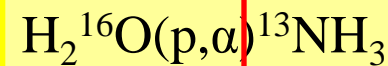
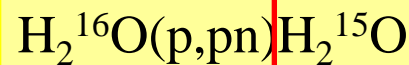
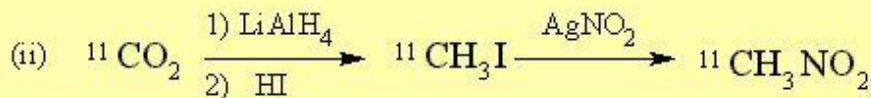
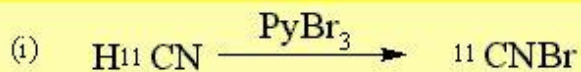
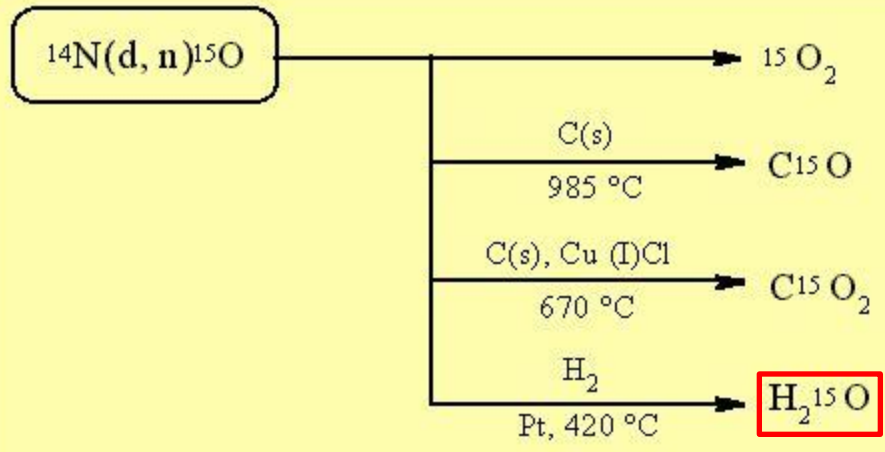
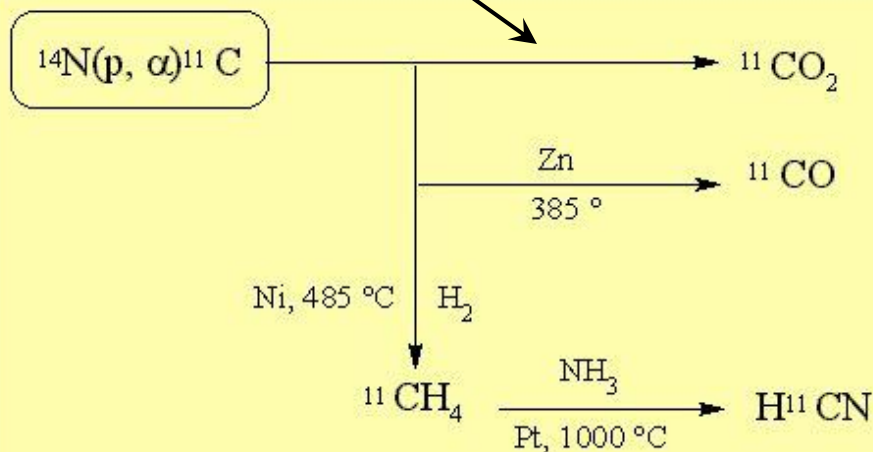
Dissolution and
electrodeposition
cylinder (front view)

CuCl_2 is not a useful tracer; must be converted into a
RADIOPHARMACEUTICAL

Main PET nuclides: Chemical forms

Chemical forms of produced radionuclides: ^{11}C , ^{15}O , ^{13}N

(Traces of oxygen)



(3-10 mM ethanol)

Real tracer

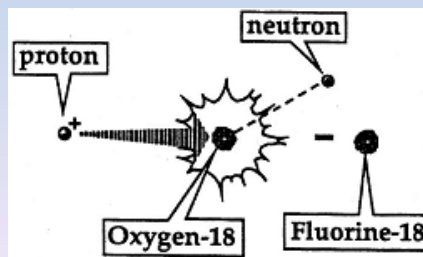
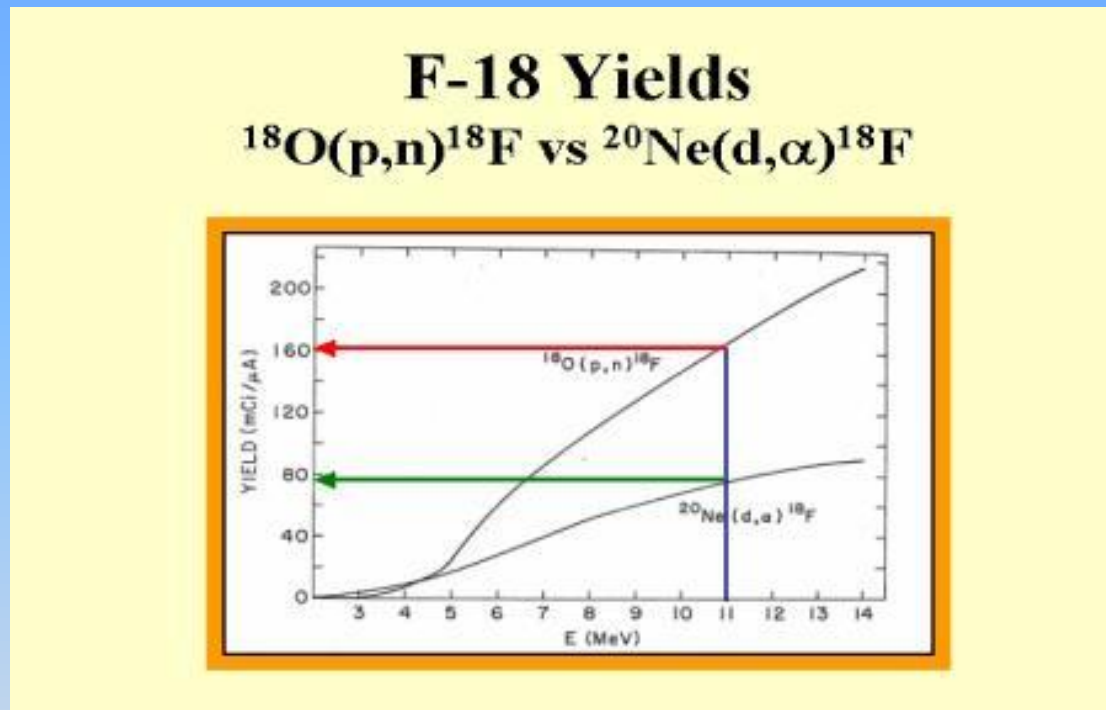
Chemical forms of produced radionuclides: ^{18}F

$[^{18}\text{F}]\text{F}_2$:

- Difficult handling
- Too high reactivity (conversion to $[^{18}\text{F}]\text{AcOF}$, $[^{18}\text{F}]\text{N}$ -fluorolactams, $[^{18}\text{F}]\text{XeF}_2$)
- Maximum rcy 50%
- Lower production rate
- c.a. production only
- Electrophilic approach

$[^{18}\text{F}]\text{HF}$:

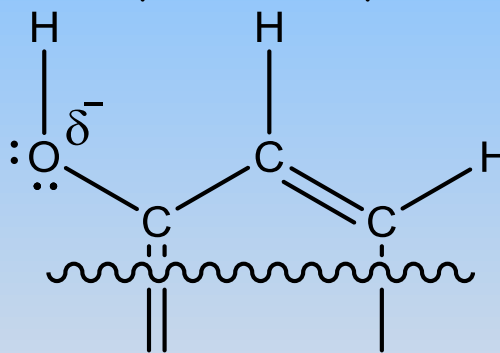
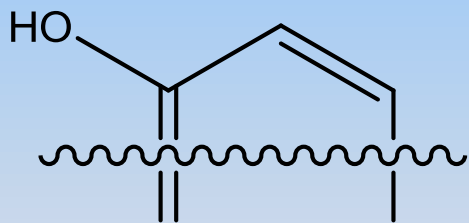
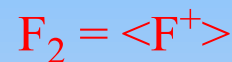
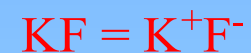
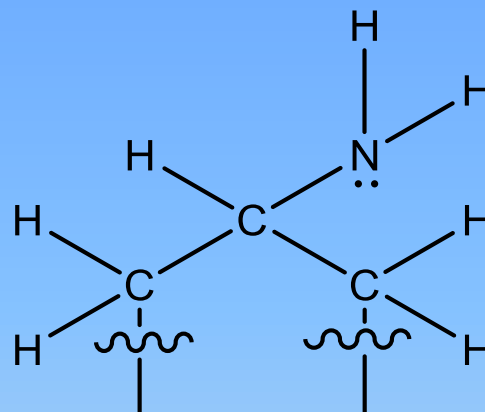
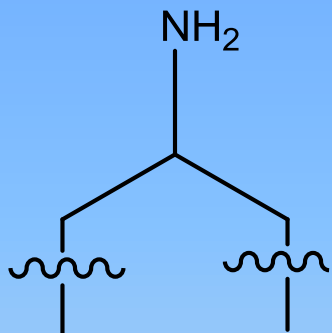
- Easy handling
- High s.a.
- Nucleophilic approach
- Low reactivity



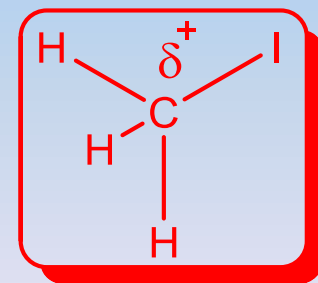
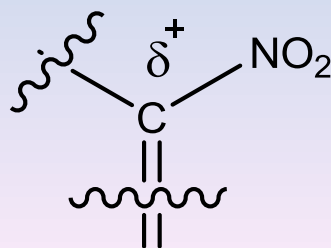
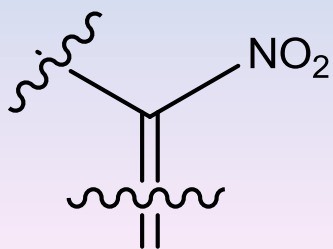
On a 13MeV GE PETrace:

- 1,5 mL $^{18}\text{O}\text{-H}_2\text{O}$ target
- 30-35 μA
- 2 hours irradiation
- 100-150 GBq

Basic organic chemistry update...

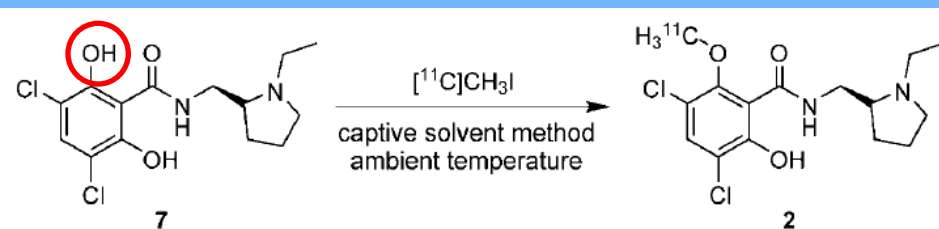


BOND
POLARIZATION
concept

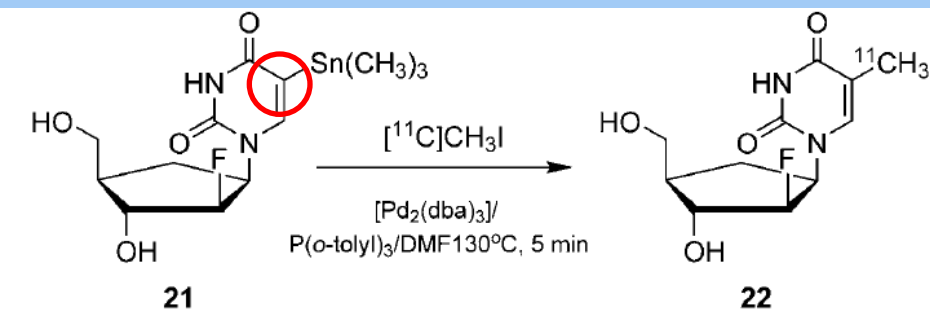


^{11}C radiochemistry

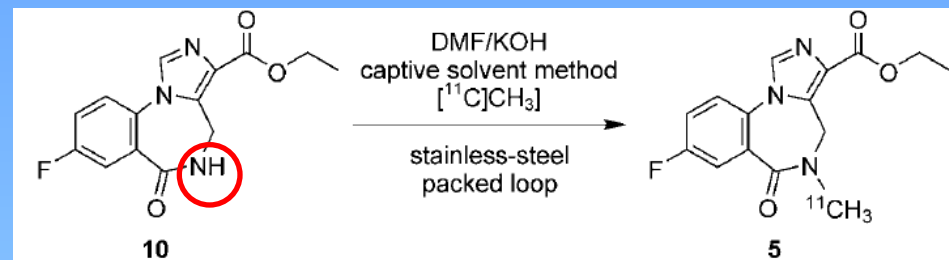
^{11}C methylation reactions



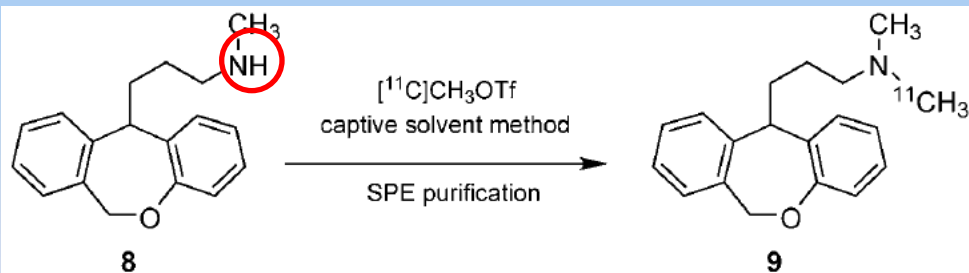
Scheme 4. O-Selective ^{11}C methylation of **7** to form $[^{11}\text{C}]$ raclopride (**2**), which is used to image dopamine D2/D3 receptors.



Scheme 13. Synthesis of $[^{11}\text{C}]$ FMAU (**22**) by palladium-mediated $[^{11}\text{C}]\text{CH}_3\text{I}$ Stille cross-coupling reactions.

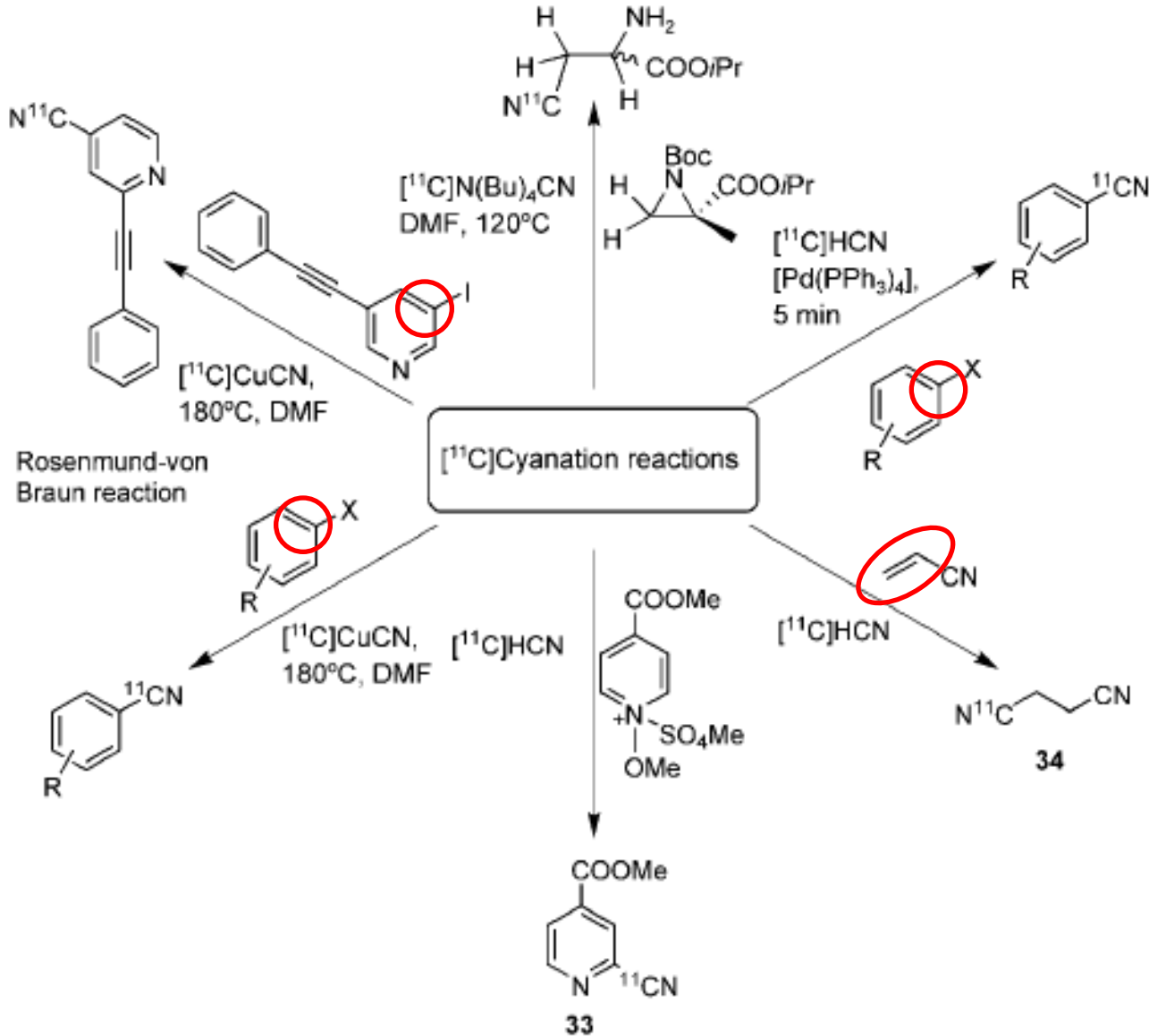


Scheme 6. N-Selective ^{11}C methylation of **10** to form $[^{11}\text{C}]$ flumazaniil (**5**), a benzodiazepine antagonist that prevents the enhancement of GABA activity.

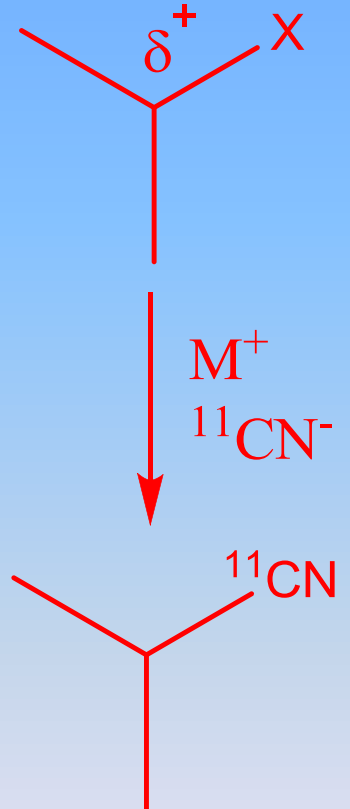


Scheme 5. N-Selective ^{11}C methylation of nordoxepin (**8**) to form $[^{11}\text{C}]$ doxepin (**9**), a histamine H1 receptor antagonist and antidepressant.

11C cyanation reactions

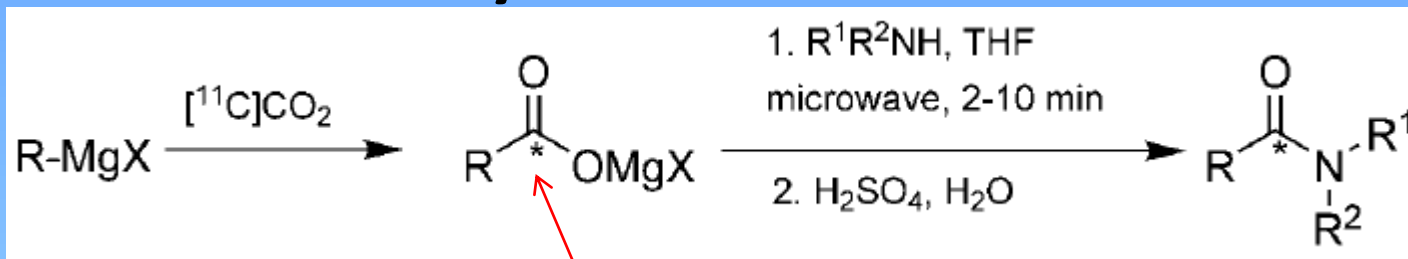


Nucleophilic substitution

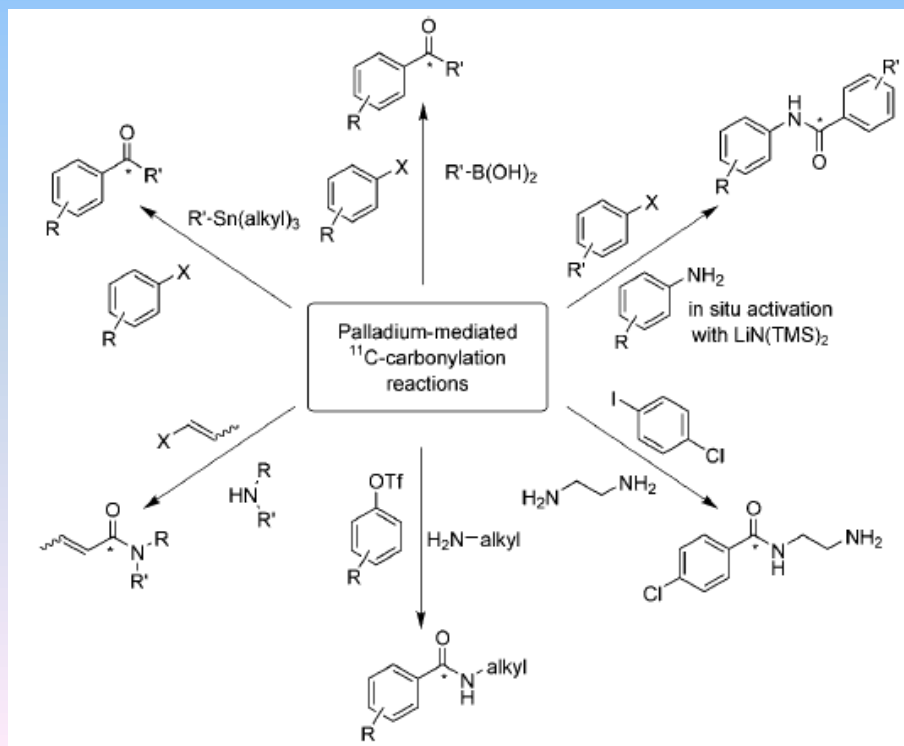


KCN? Poisonous?
NO!! (s.a.)

11C carbonylation and carboxylation reactions



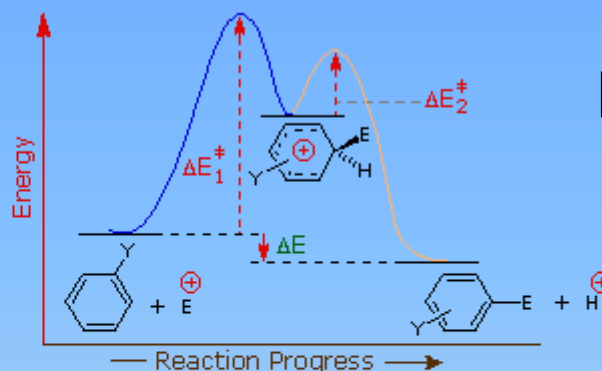
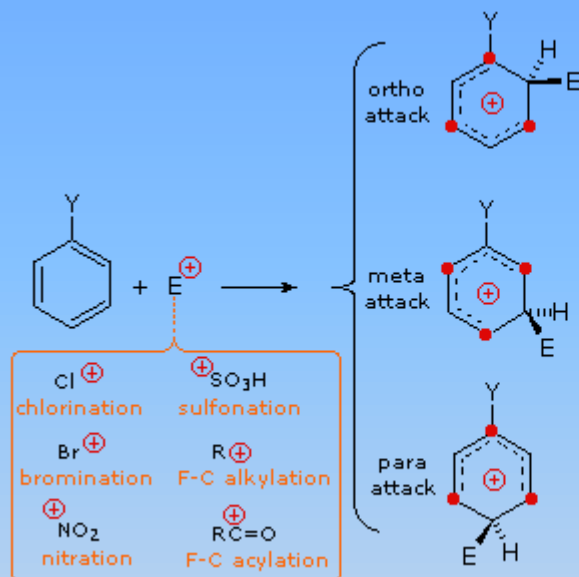
Simple route to 11C-acetic acid and 11C-fatty acids



Carbonylation =
Insertion of a C=O
group between
two carbon atoms

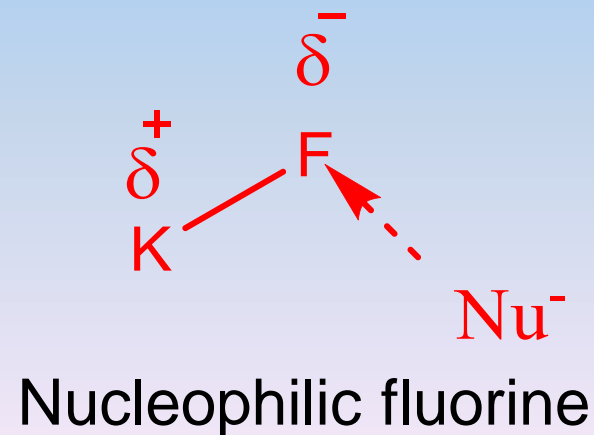
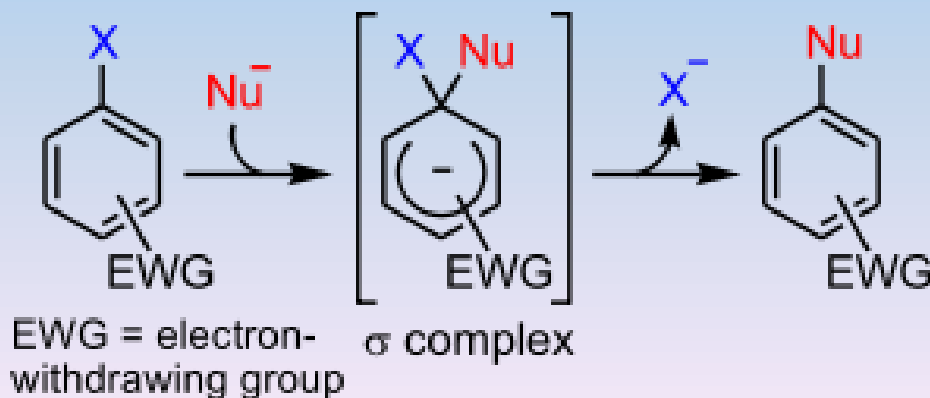
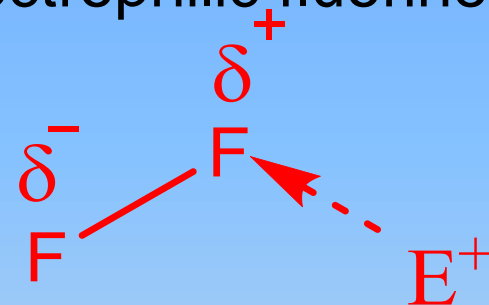
^{18}F radiochemistry

Electrophilic and nucleophilic aromatic substitution

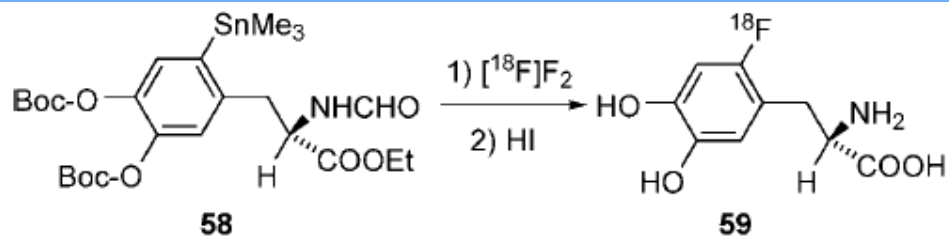


The first step in electrophilic aromatic substitution (shown here) is both rate-determining and product determining. To see the effect various substituents (-Y) have on the orientation of such substitutions press one of the buttons provided below.

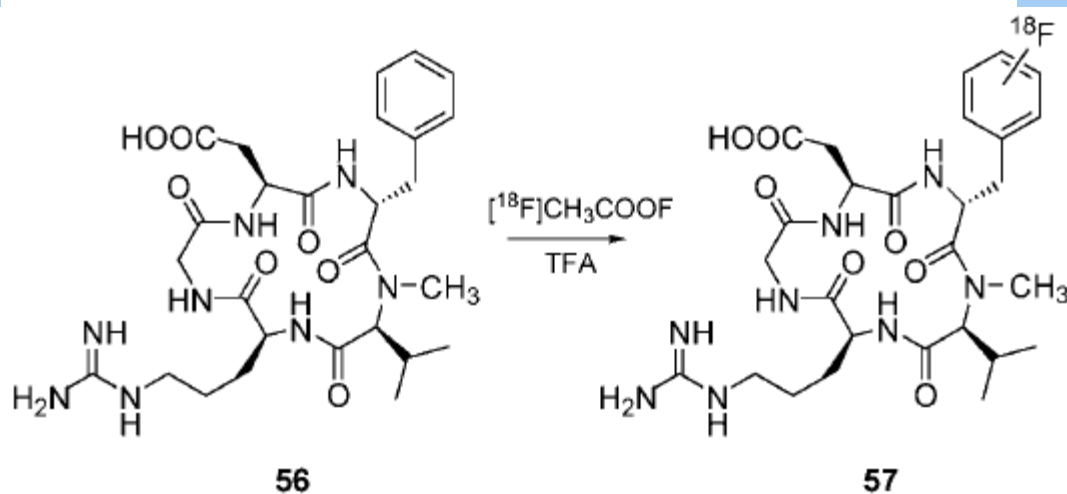
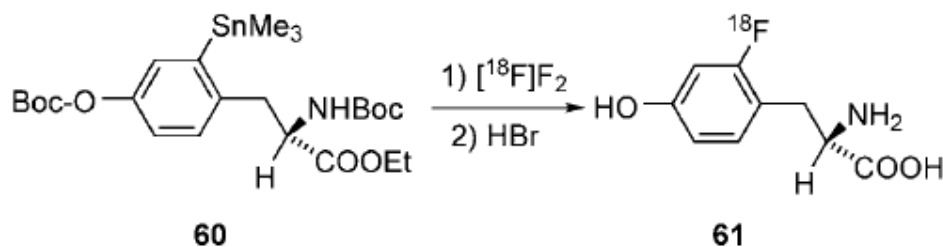
Electrophilic fluorine



^{18}F electrophilic reactions



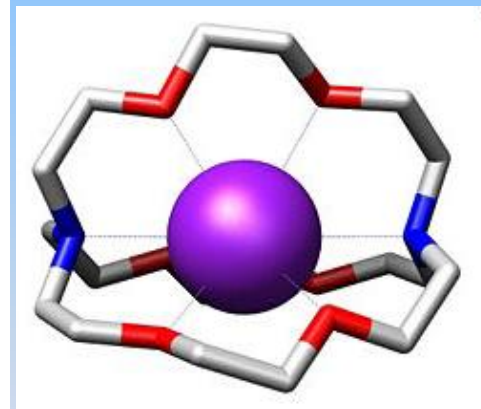
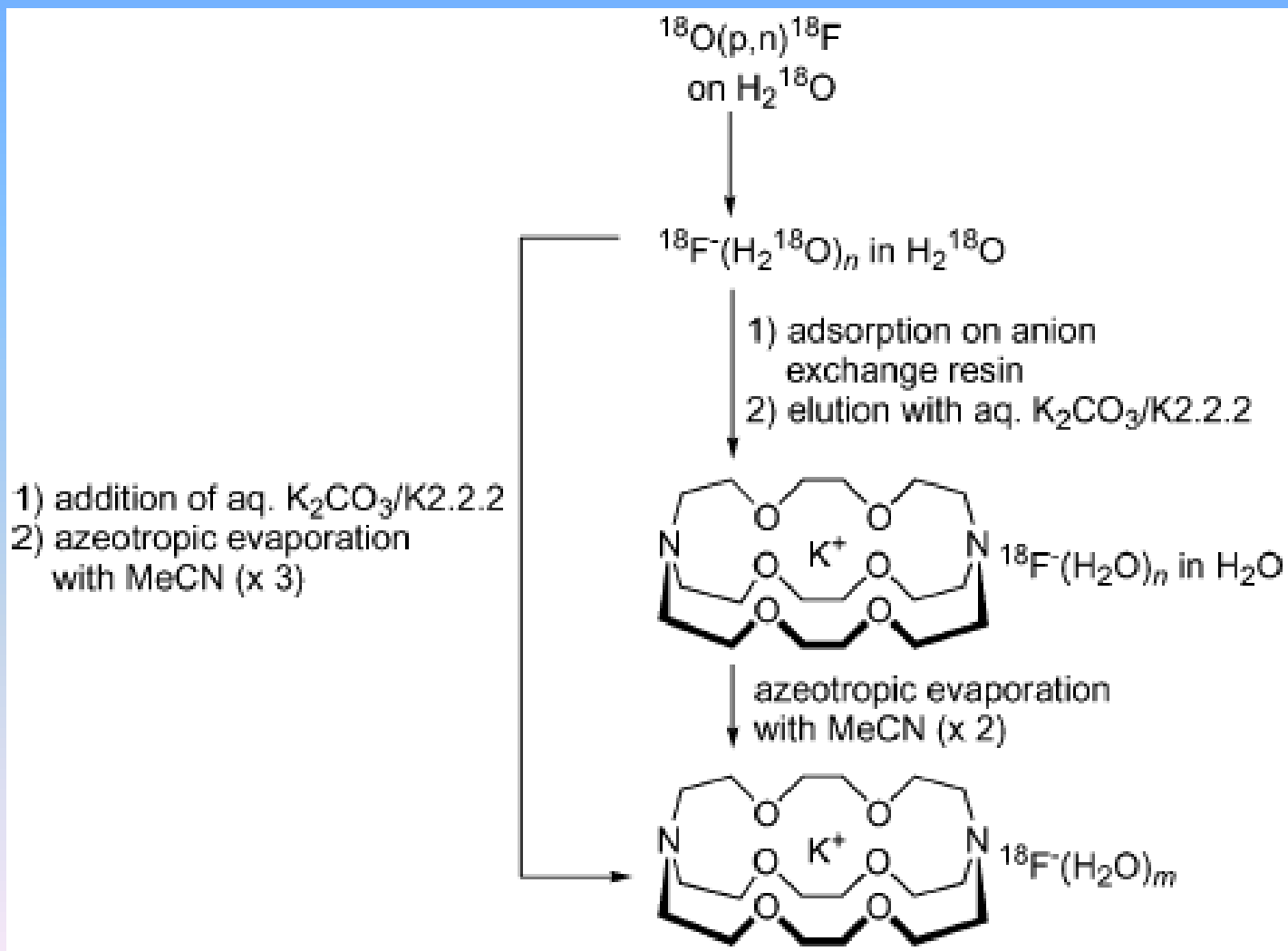
F_2 is a very reactive reagent (needs carrier added conditions in target) and its direct use must be regioselective by use of organometallics



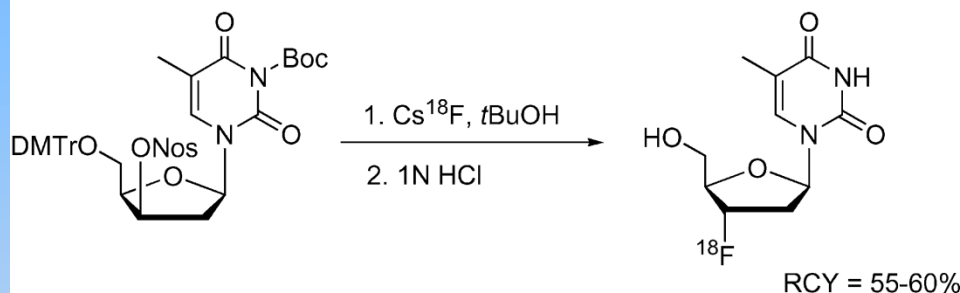
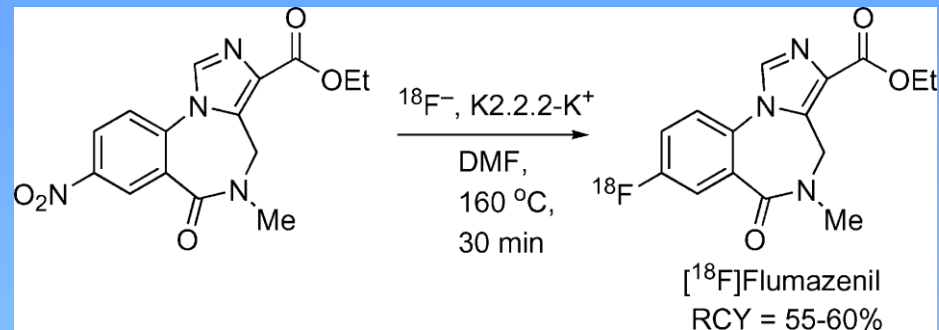
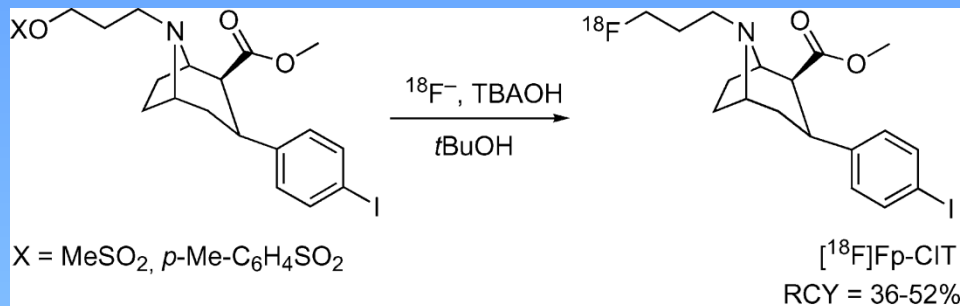
F_2 activity can be “tamed” by generating milder fluorinating species, such as acetyl fluoride or XeF_2

Scheme 33. Electrophilic ^{18}F fluorination of cyclic RGD peptides using $[^{18}\text{F}]\text{CH}_3\text{COOF}$.

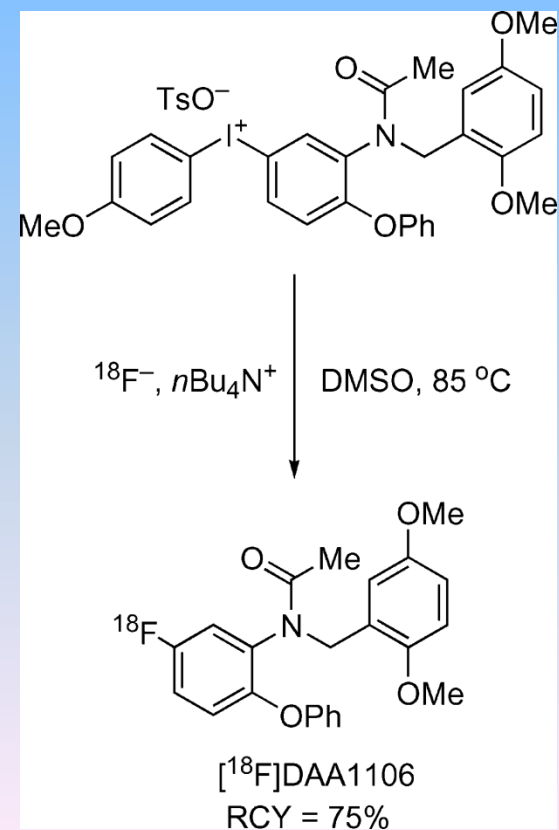
18F nucleophilic reactions: activation step



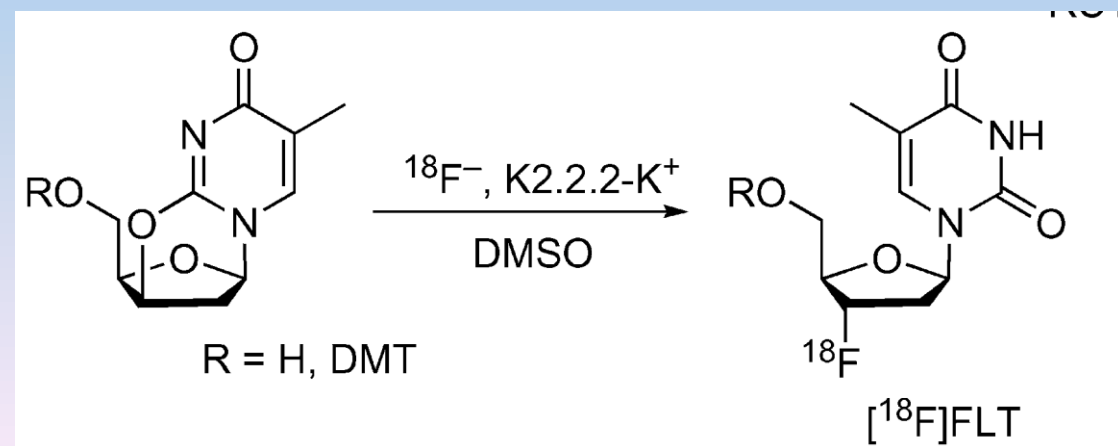
18F nucleophilic reactions: direct



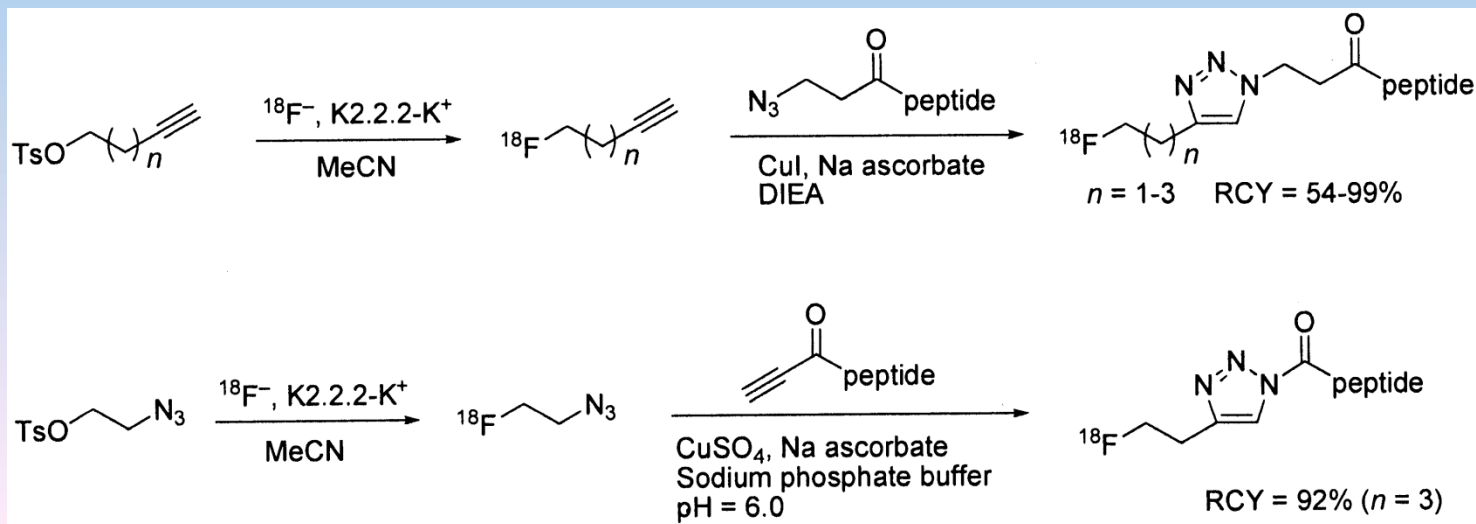
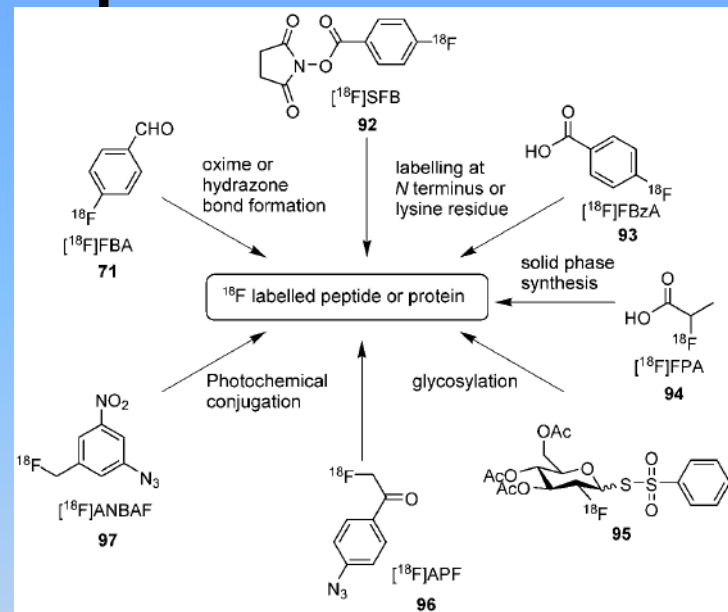
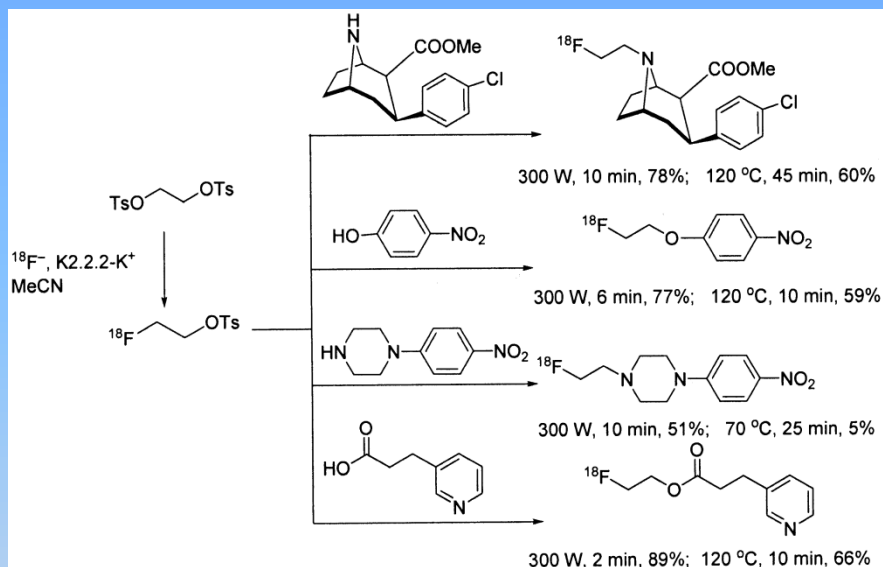
Aromatic substitution



Aliphatic substitution



18F nucleophilic reactions: prosthetic groups use



Specific activity

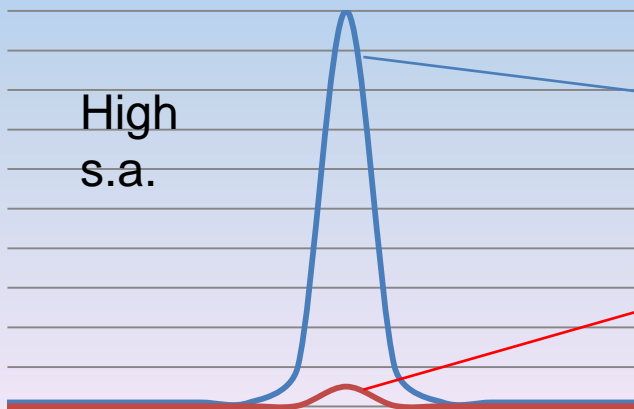
Nuclide	Maximum SA		Practical SA
	mCi μg^{-1}	Ci μmole^{-1}	Ci μmole^{-1}
^{11}C	838,000	9,220	<100
^{18}F	95,000	1,170	10–20 as F^- < 0.03 as F_2
^{68}Ga	40,600	2,766	
^{67}Ga	597	47	<3.35
^{111}In	423	40	<5.55
^{123}I	1,926	237	
^{124}I	250	31	

Carrier added (c.a.):

The correspondent stable nuclide is added to the reaction/preparation

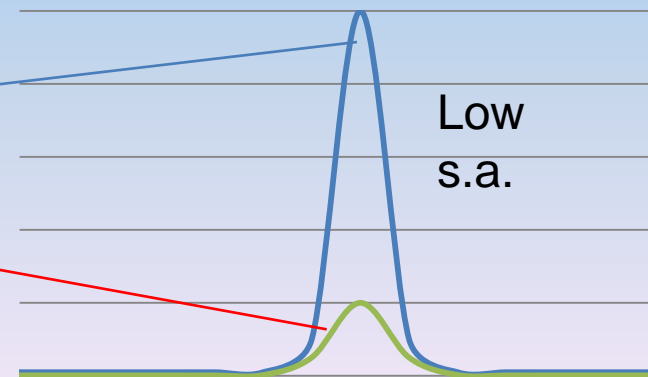
No carrier added (n.c.a.): no addition of carrier

Carrier free (c.f.): no stable isotope is present in preparation



$$s.a. = \frac{\text{activity}_{trac}}{\text{mass}_{trac}}$$

Quantify the chemical entity generating the activity



Automated synthesis systems

Need for automation

GE, Siemens

Eckert&Ziegler, Raytest, IBA, Scintomics, Advion



??



50 x 40 x 50 cm

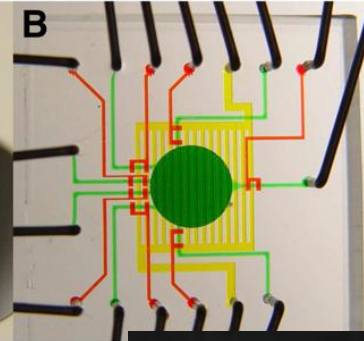
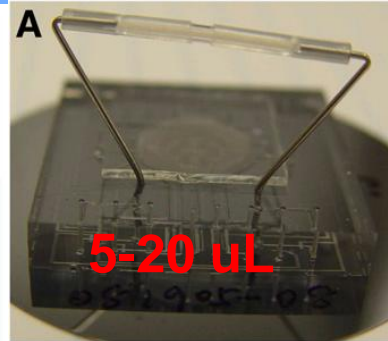
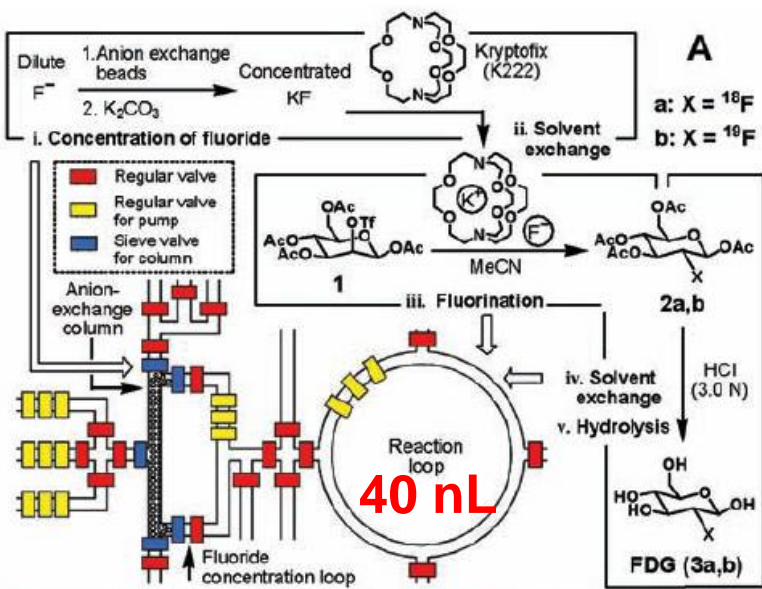
30 x 25 x 50 cm

30 x 20 x 20 cm

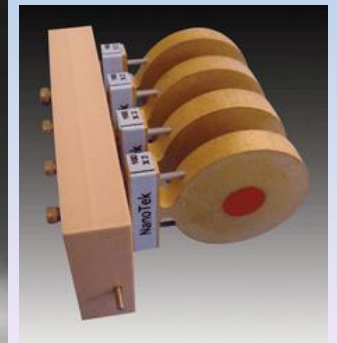
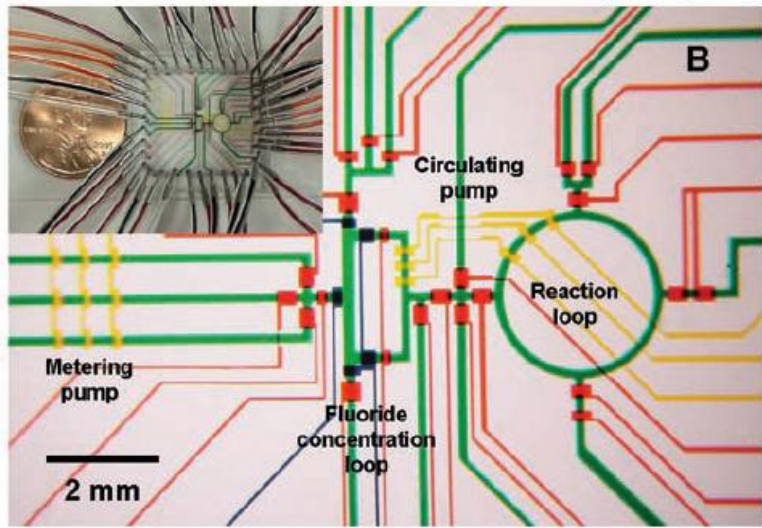
Need for automation: price...



New avenues for automation



Microfluidics



Radiopharmaceuticals: general



GeorgedeHevesy

***The Father of
Nuclear Medicine:***

George de Hevesy
(1885 - 1966)

**Received Nobel Prize
in 1943 for his
pioneering work with
isotopes as tracers.**

**Winner of Atom for
Peace Award 1959.**

The first practical application of radioisotopes



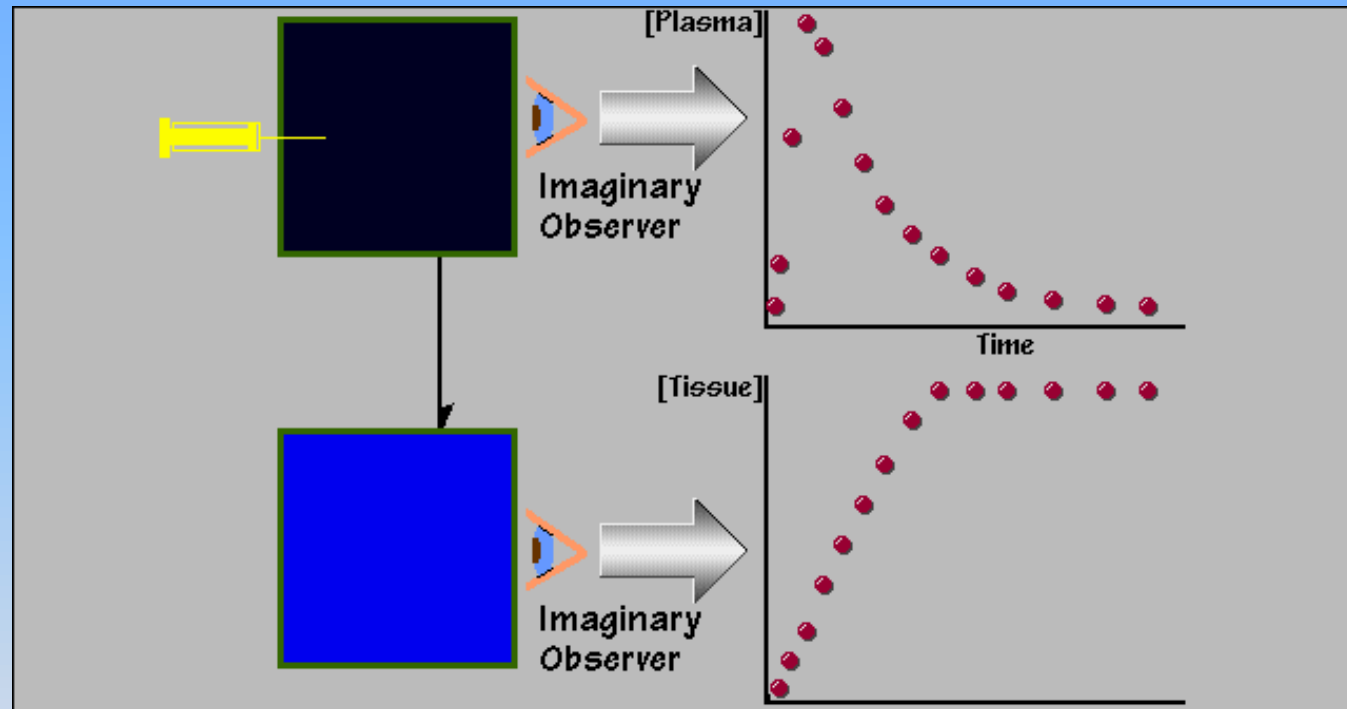
George de Hevesy & his landlady:

- **Using radioactive material he proved two things:**
 - The landlady was indeed "recycling" leftovers from their plates!
 - More importantly, that small amounts of radioactive materials could be used to "trace" the fate of a substance in a system.

Biodistribution generalities

Target/non-target ratio

Blood extraction of the radiopharmaceutical to accumulate selectively on target organs

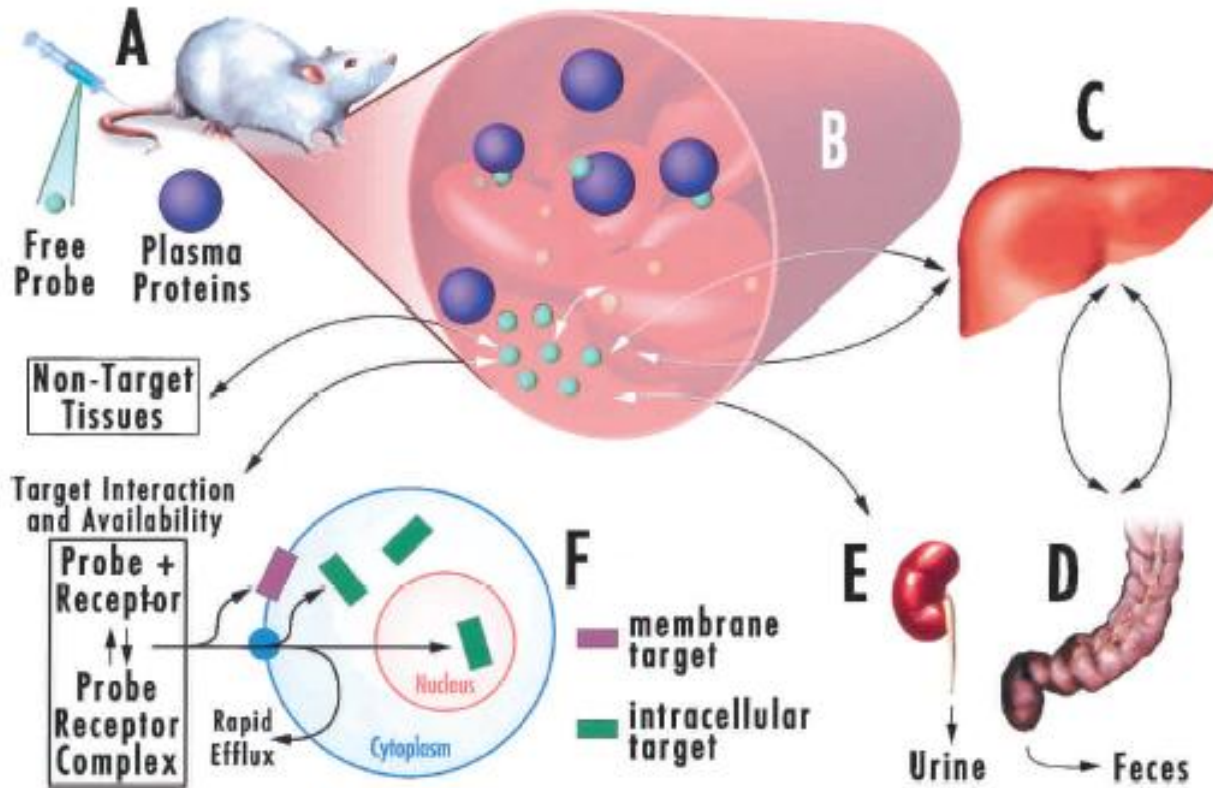


Factors

- Chemical nature of tracer
- Blood flow
- Membrane permeation
- Competition with endogenous ligands

Simple 2 compartment model

A more complete scheme...

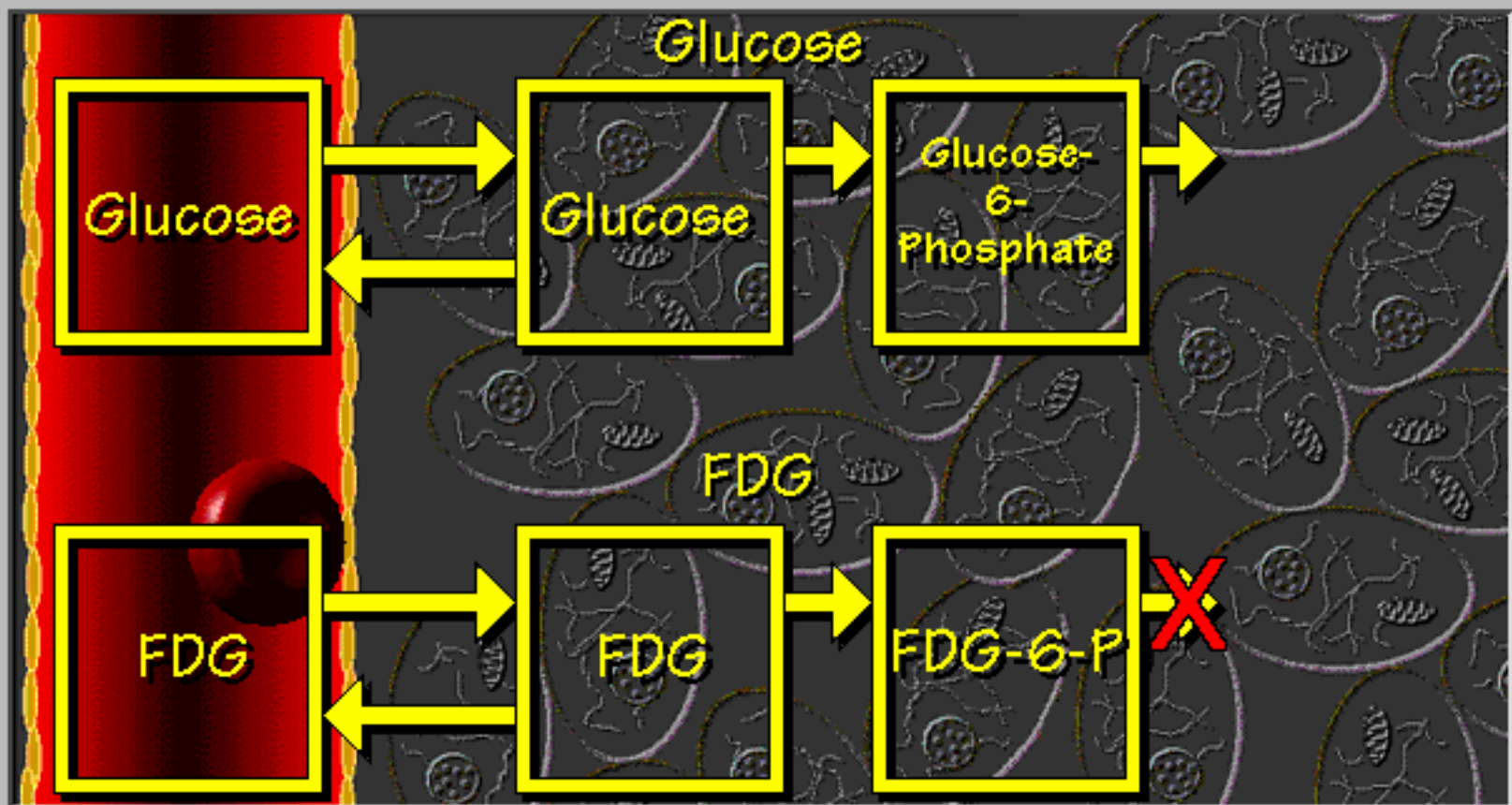


Pharmacokinetics of molecular imaging probes. Molecular imaging probes need to overcome many biological barriers when administered to living subjects. These probes are subject to all the pharmacokinetic rules and constraints that govern the concentration of "drugs" in plasma, including absorption/delivery (A), distribution (B), metabolism (C), excretion/reabsorption in the enterohepatic circulation (D), urinary excretion (E), and other factors within the vascular compartment (B; e.g., plasma half-life, protein binding). Rapid excretion, nonspecific binding/trapping in nontarget tissues, metabolism, and delivery barriers are all important obstacles to be overcome before availability to target(s) for interaction (F).

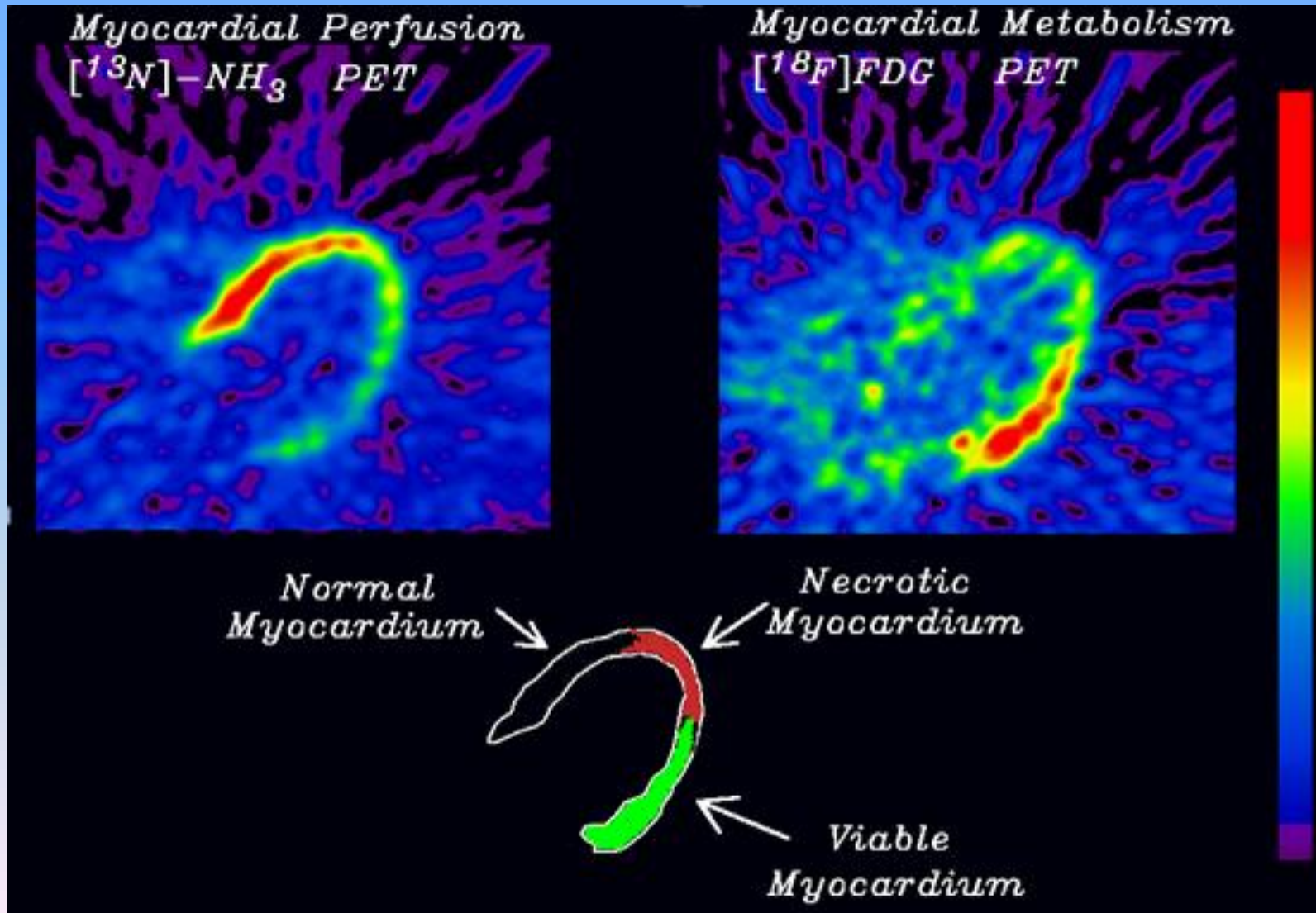
A: Administration
D: Distribution
M: Metabolism
E: Excretion
T: Toxicology

A
D
M
E
T

[¹⁸F]FDG model



Classical example: ^{13}N -NH₃ for perfusion, ^{18}F -FDG for metabolism

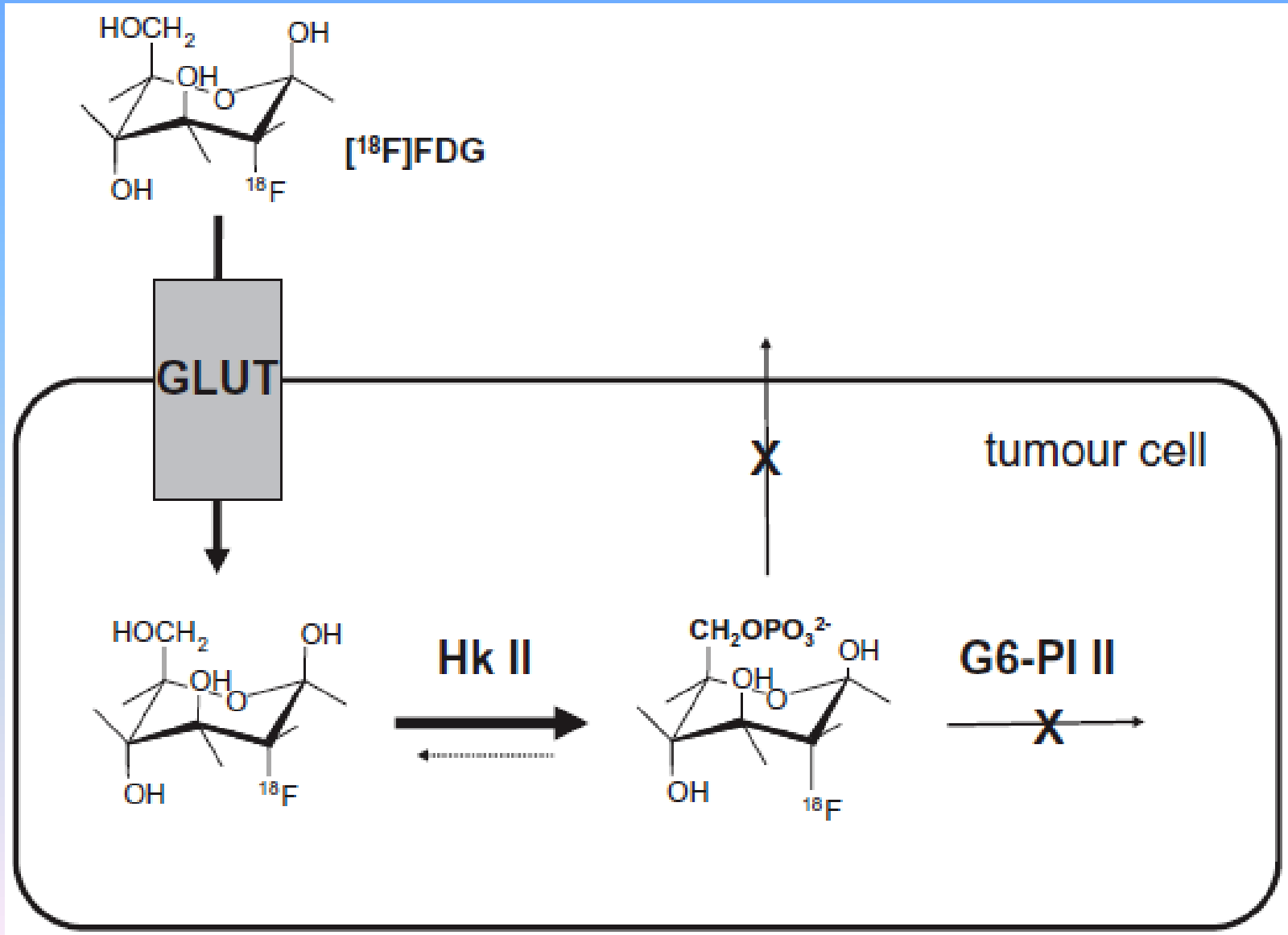


Working principles of Selected radiotracers

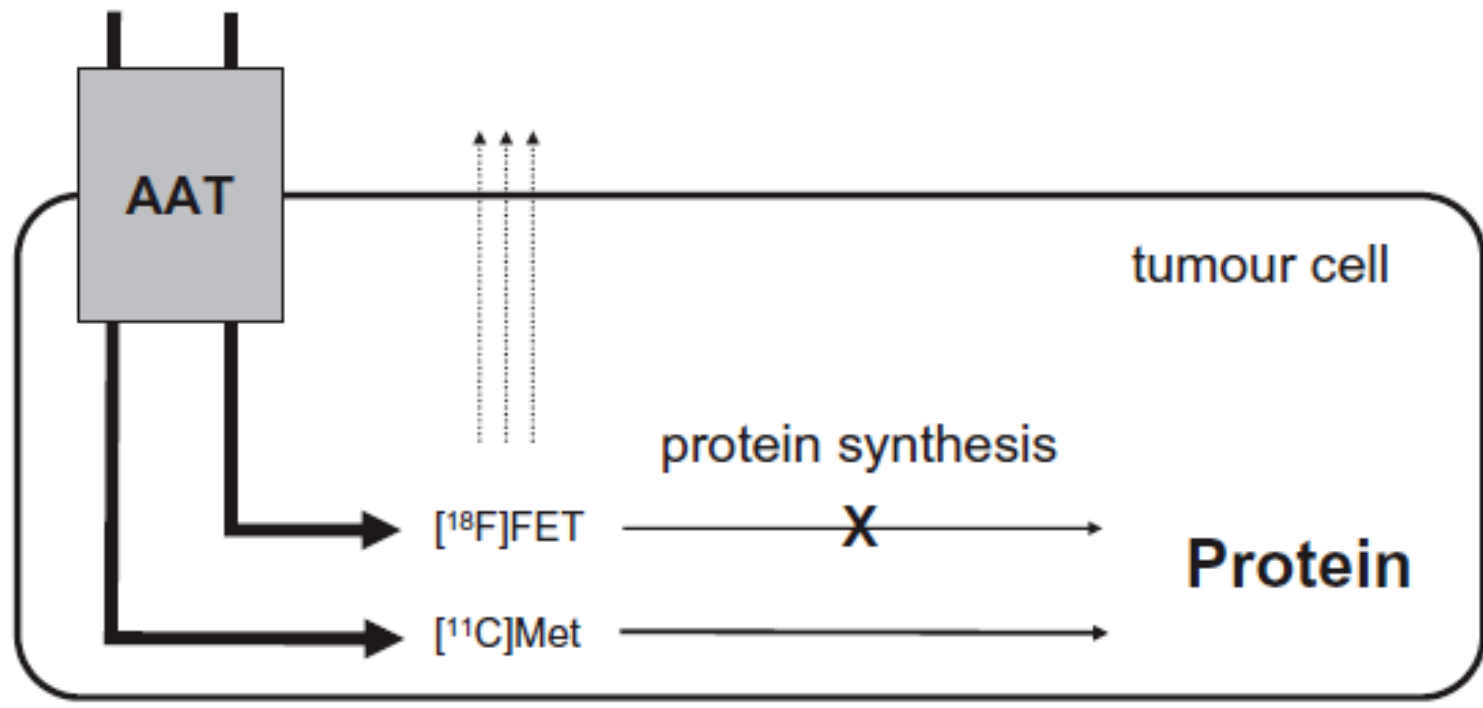
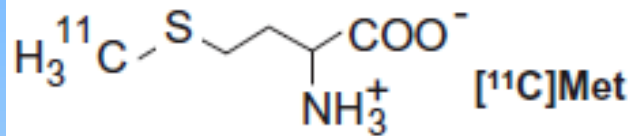
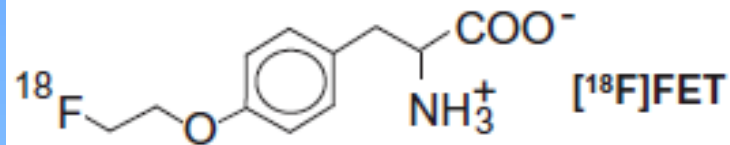
PET radiopharmaceuticals

Molecular uptake mechanism	Tracer	Isotope	Organs of highest physiological uptake ^a	Availability
Amino acid transport and protein synthesis	Methionine	C-11	Liver, salivary glands, lachrymal glands, bone marrow, pancreas, bowels, renal cortical, urinary bladder	In-house production/cyclotron
	Fluoroethyltyrosine	F-18	Pancreas, kidneys, liver, heart, brain, colon, muscle	In-house production/cyclotron ^b
	FDOPA	F-18	Pancreas, liver, duodenum, kidneys, gallbladder, biliary duct	Commercially available
Glucose metabolism	FDG	F-18	Brain, myocardium, breast, liver, spleen stomach, intestine, kidney, urinary bladder, skeletal muscle, lymphatic tissue, bone marrow, salivary glands, thymus, uterus, ovaries, testicle, brown fat	Commercially available
Proliferation	FLT	F-18	Bone marrow, intestine, kidneys, urinary bladder, liver	In-house production/cyclotron ^b
Hypoxia	FMISO	F-18	Liver, urinary excretion	In-house production/cyclotron ^b
	FAZA	F-18	Kidneys, gallbladder, liver, colon	In-house production/cyclotron
	Cu-ATSM	Cu-64	Liver, kidneys, spleen, gallbladder ^c	In-house production/cyclotron ^b
Lipid metabolism	Choline	C-11	Liver, pancreas, spleen, salivary glands, lachrymal glands, renal excretion, bone marrow, intestine	In-house production/cyclotron
	Fluoroethylcholine	F-18	Liver, kidneys, salivary glands, urinary bladder, bone marrow, spleen	In-house production/cyclotron ^b
	Acetate	C-11	Gastrointestinal tract, prostate, bone marrow, kidneys, liver, spleen, pancreas	In-house production/cyclotron
Angiogenesis/integrin binding	Galacto-RGD	F-18	Bladder, kidneys, spleen, liver	In-house production/cyclotron
	AH111585	F-18	Bladder, liver, intestine, kidneys	In-house production/cyclotron
SSTR binding	DOTATOC	Ga-68	Pituitary and adrenal glands, pancreas, spleen, urinary bladder, liver, thyroid	In-house production/generator
	DOTATATE	Ga-68	Spleen, urinary bladder, liver	In-house production/generator

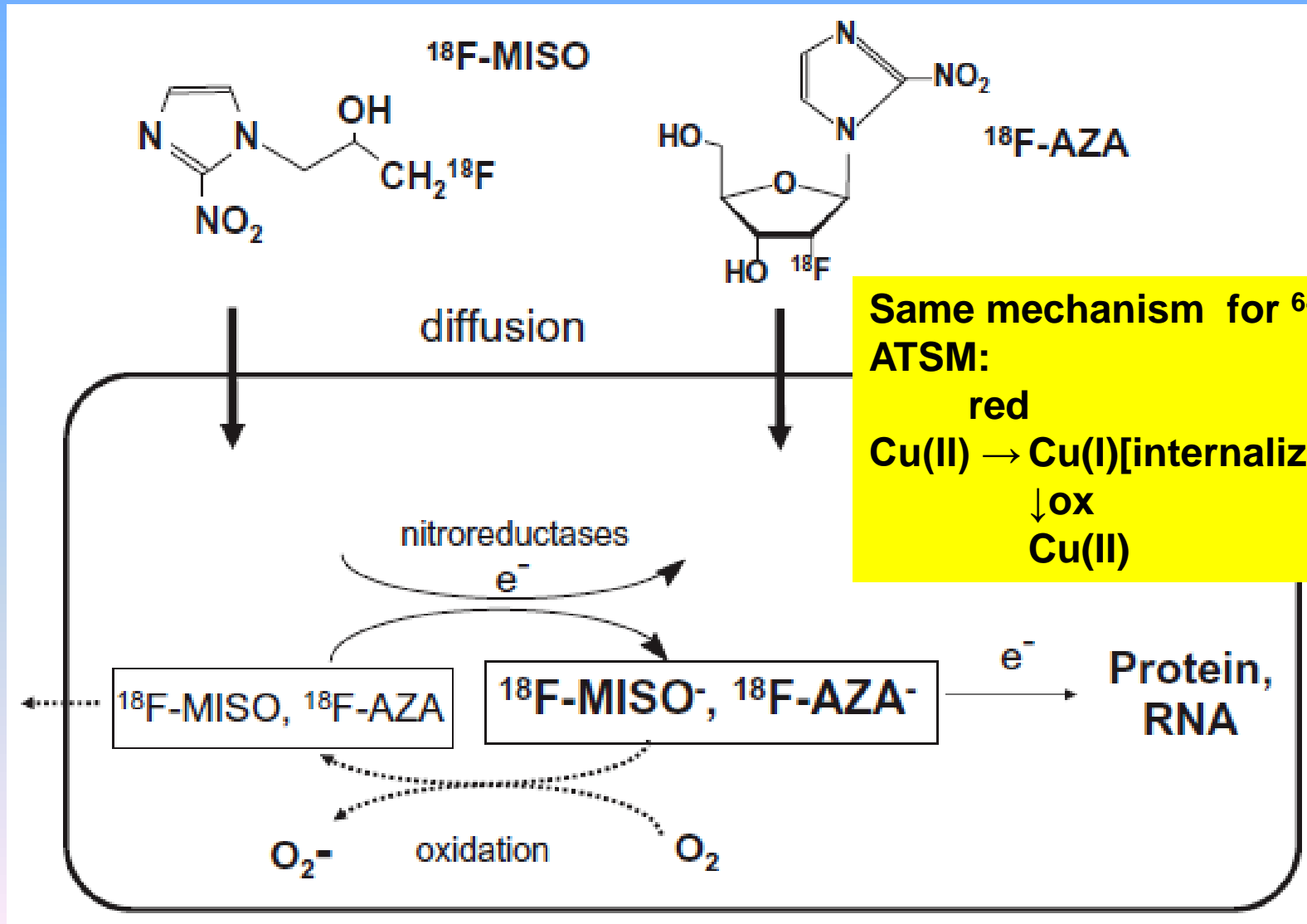
Working principle: FDG



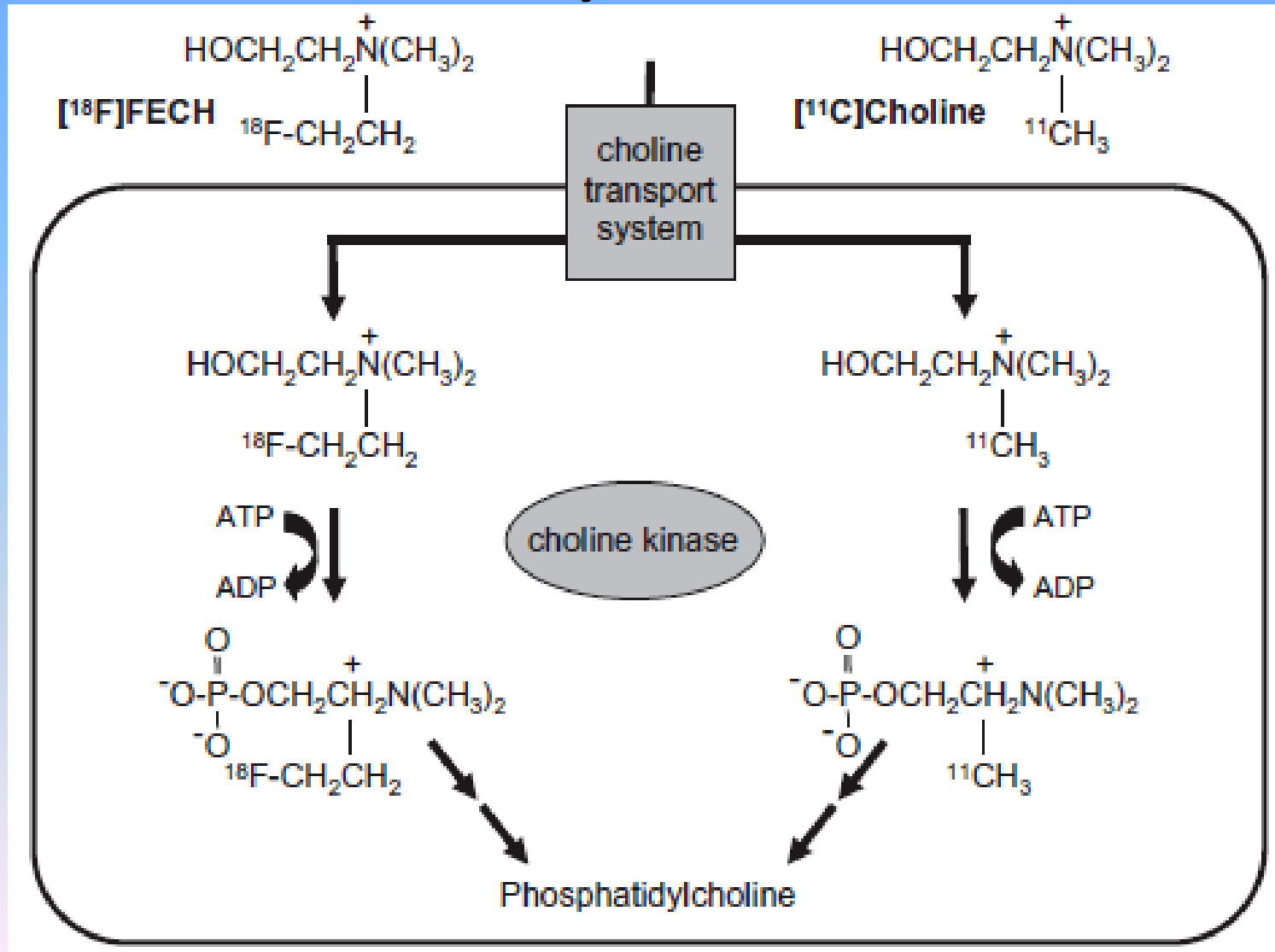
Working principle: labelled amino acids



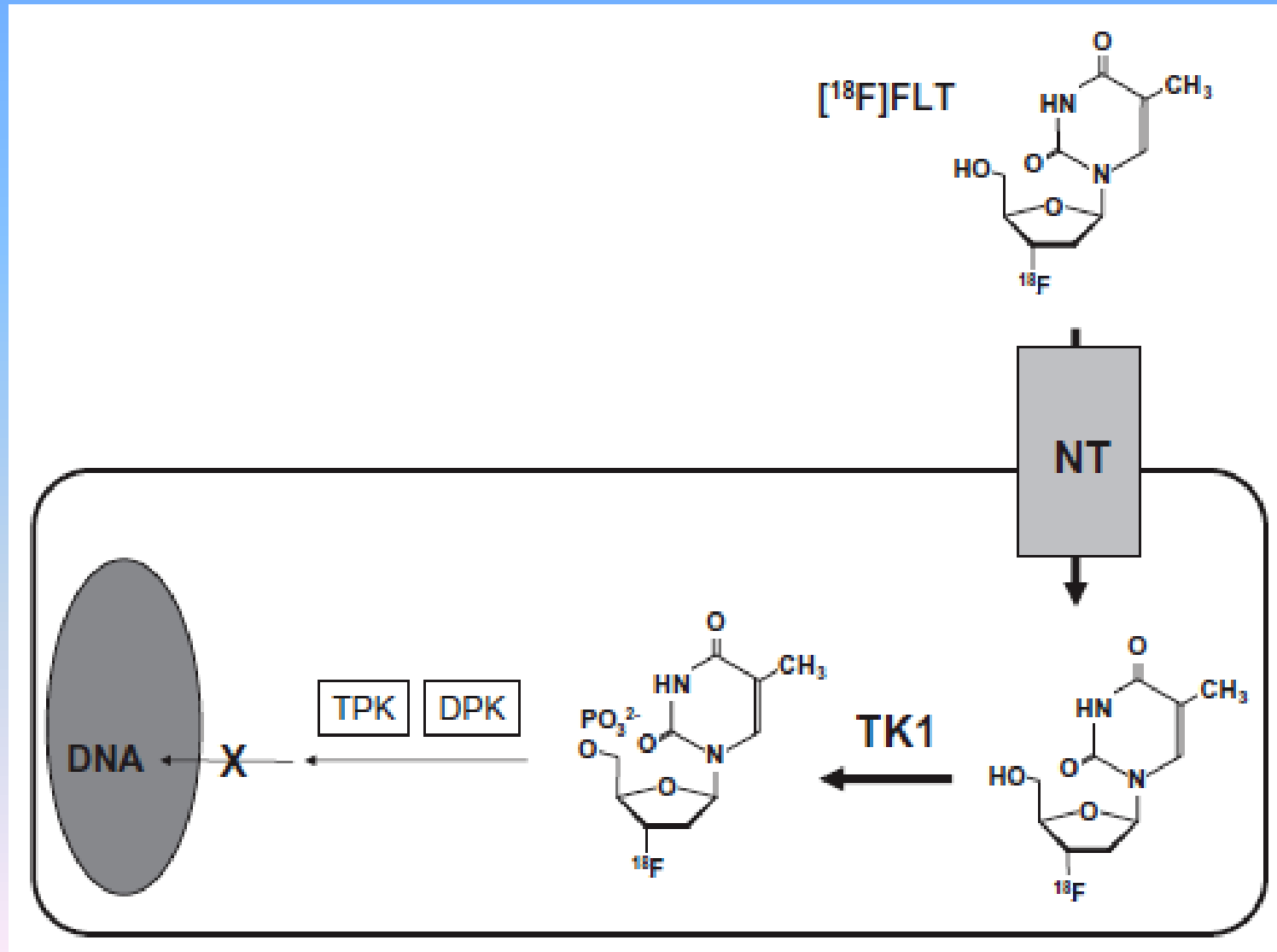
Working principle: hypoxia agents



Working principle: membrane biosynthesis



Working principle: labelled nucleosides



Working principle: angiogenesis and somatostatin tracers

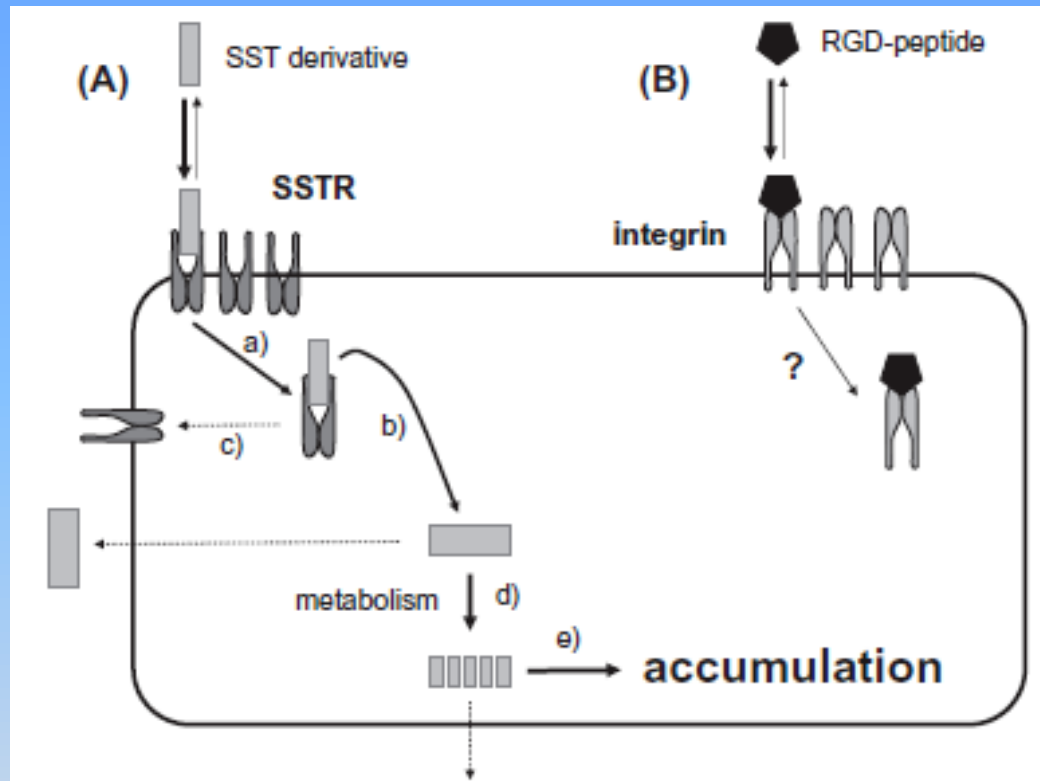
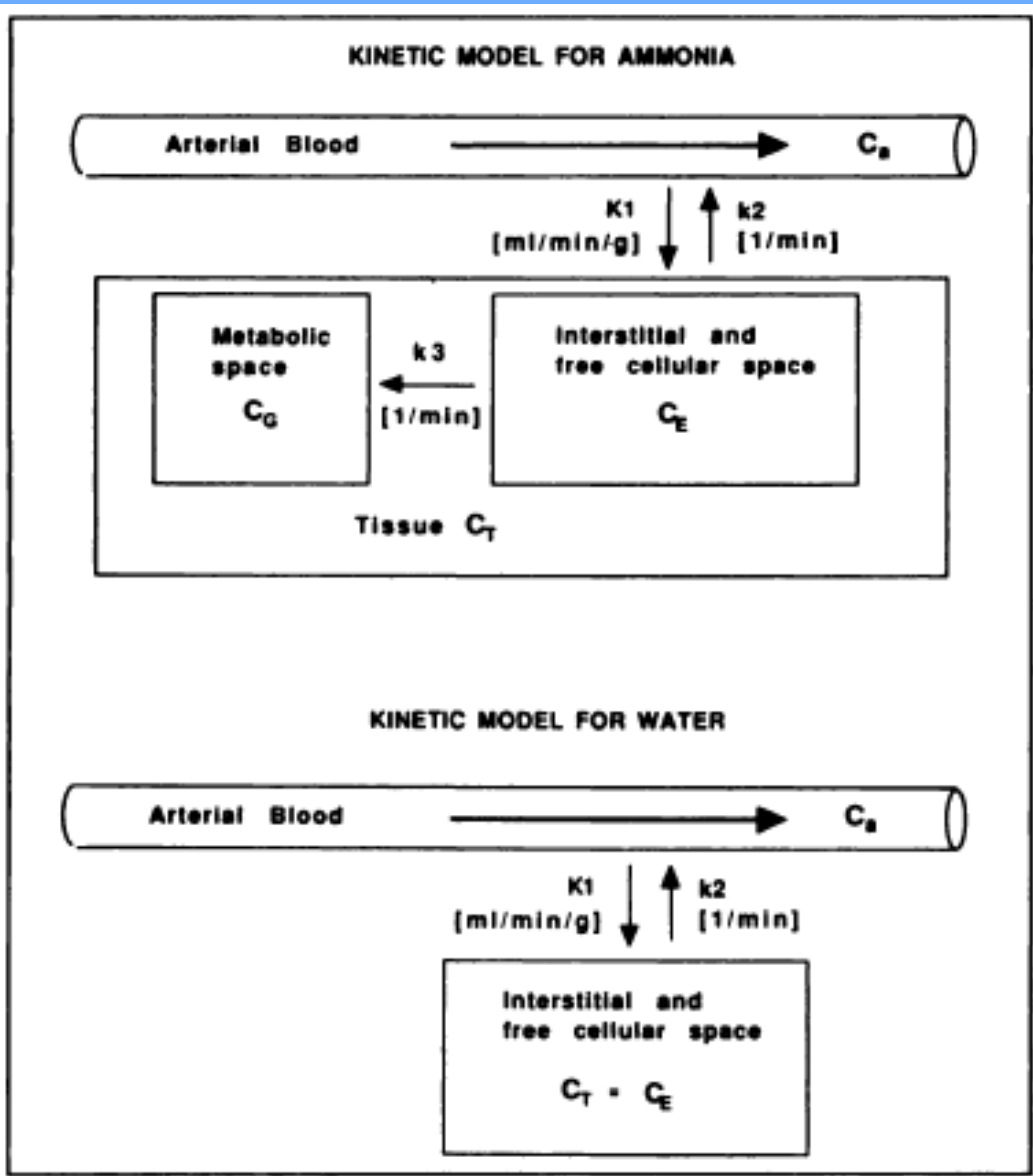


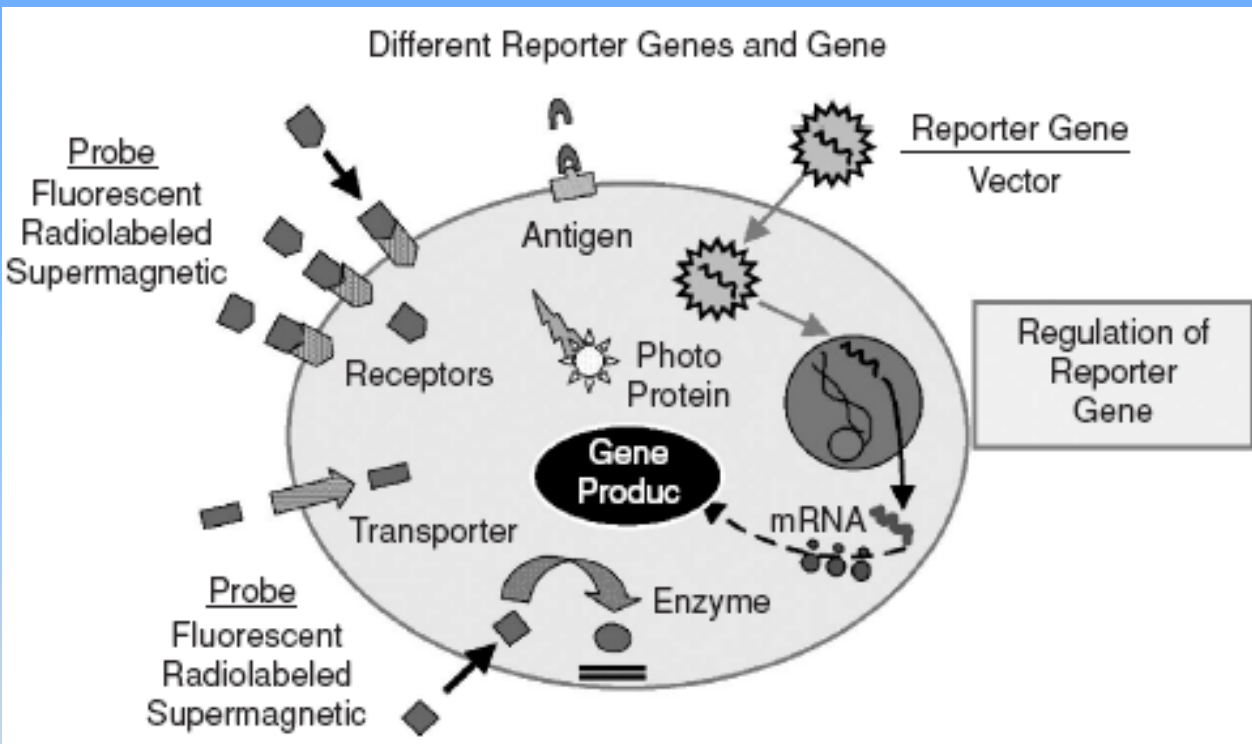
Fig. 6. Schematic presentation of uptake of radiolabelled-somatostatin derivatives and radiolabelled-RGD-peptides: (A) Somatostatin derivatives (e.g. radiolabelled DOTATOC and DOTATATE) bind to the somatostatin receptor (SSTR) and are internalized via endocytosis (a). After release of the tracer (b) the receptor can be recycled (c). The major amount of the radiolabelled somatostatin derivative is metabolized (d) resulting in fragments which can not penetrate the cell membrane (e). (B) Radiolabelled RGD-peptides bind with high affinity to the integrin (especially $\alpha v \beta 3$). It is discussed that the receptor ligand complex can also be internalized. However, at the moment corresponding studies are missing. In *in vitro* internalization assays only small amounts of activity are found inside the cell (see e.g. Ref. [69]). Thus, it is unclear if *in vivo* accumulation is mainly based on receptor-binding or internalization.

Tracer model for $^{15}\text{O}-\text{H}_2\text{O}$ and $^{13}\text{N}-\text{NH}_3$



Multiparametrical analysis allows to assess exactly the blood volume

Tracers for gene therapies

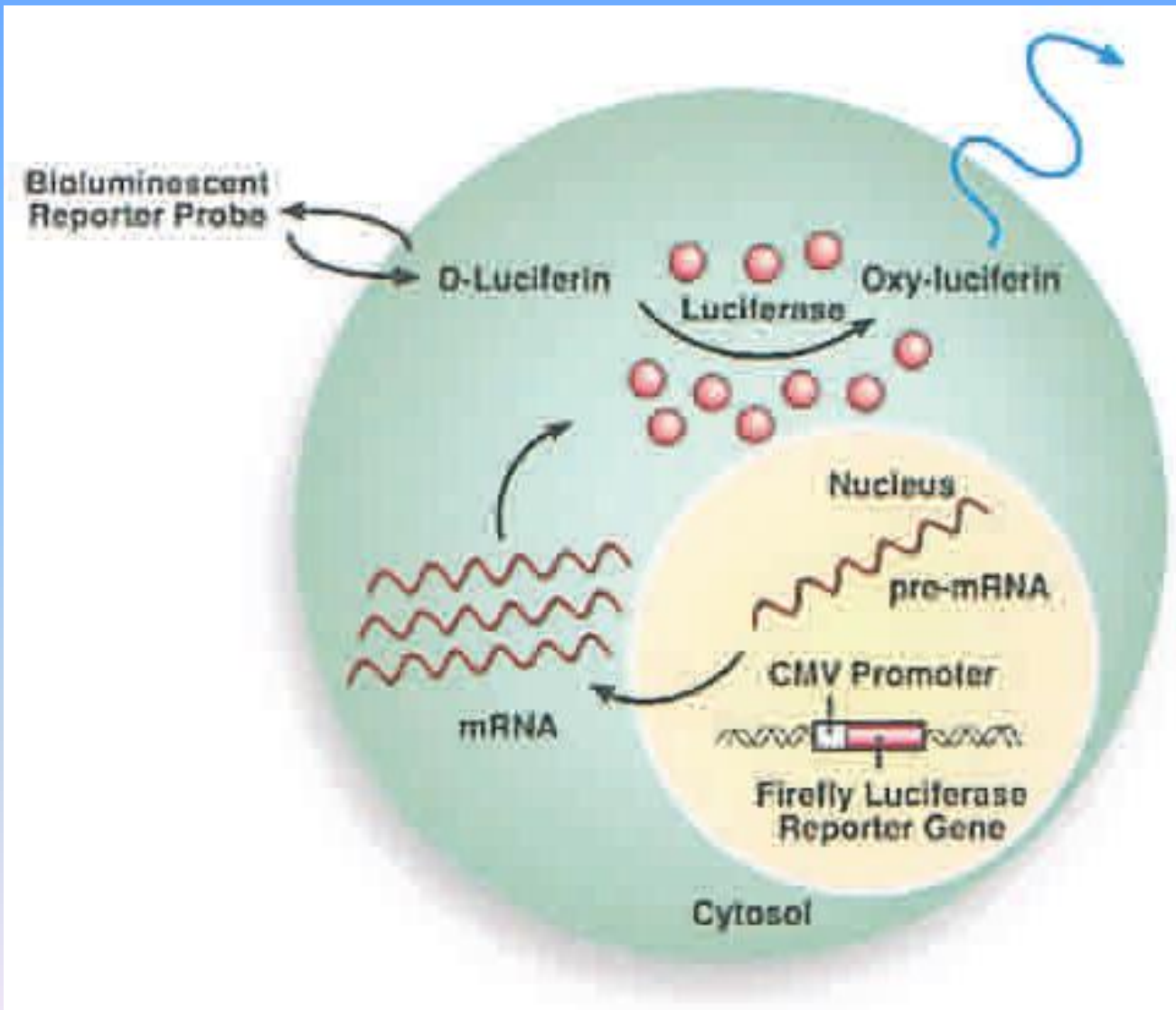


- A gene coexpresses curative protein/enzyme and a secondary agent (enzyme, receptor, protein)
- The radiotracer has a binding selectivity for the secondary agent
- Radioactive uptake will give information on the main expression

- Modify genome of a given cell population for study or curative reasons
- Problem: quantify gene expression, that is evaluation of *location, magnitude* and *persistence*

Reporter gene technique uses a small, selected set of secondary agents to be coupled with virtually any kind of curative gene

Working principle of Luciferase system

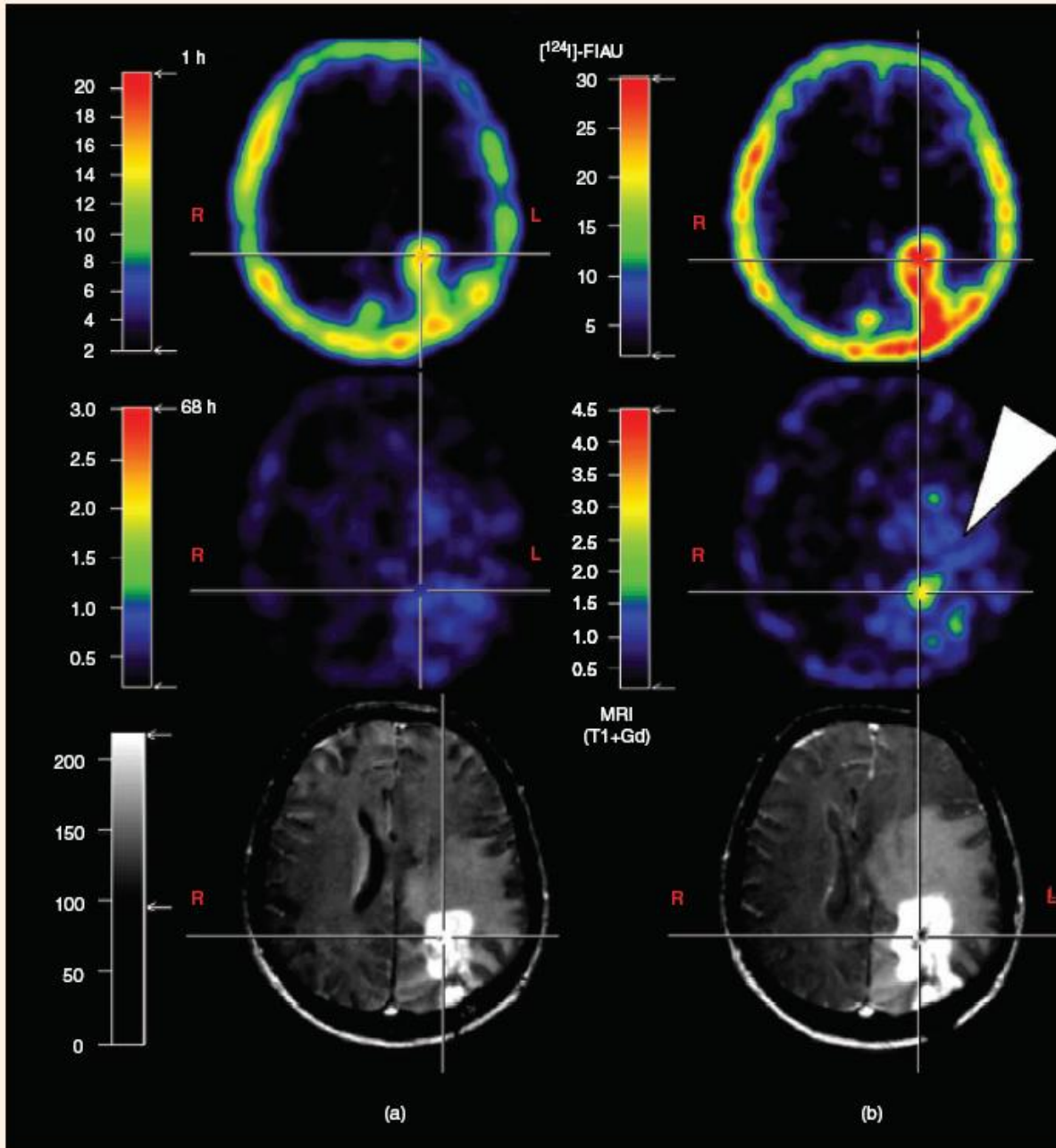


Luciferase = secondary agent

Luciferin (or luciferin analogues) = reporter gene tracer

Before
LIPO-HSV-1-*tk*
Infusion

After
LIPO-HSV-1-*tk*
Infusion



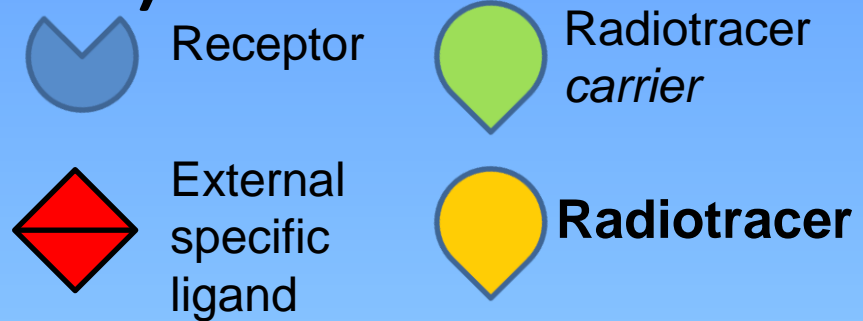
(a)

(b)

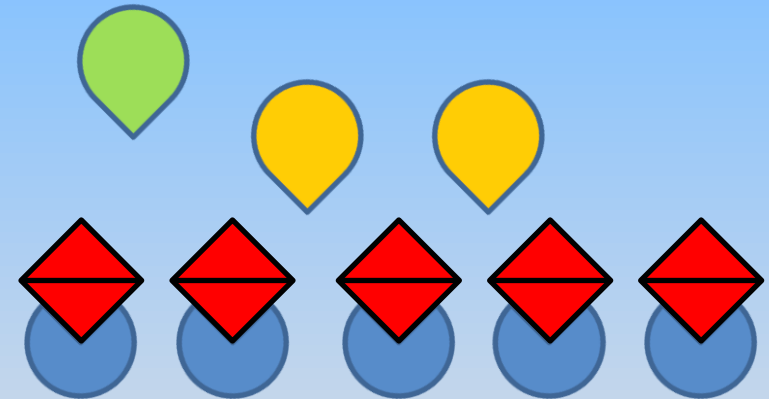
Specific binding assessment (brain receptors)



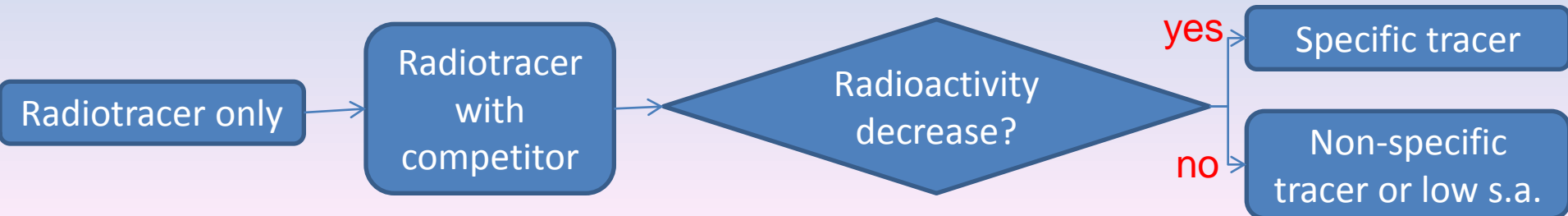
Radiotracer only, high s.a.



Radiotracer only, low s.a.



Radiotracer with competition



Concluding remarks

Testing of new radiopharmaceuticals

In vitro

- Cell culture of target tissue

Ex vivo

- Binding on tissue slices
- Biodistribution on excised organs

In vivo

- “Simulated” animal model
- Bioengineered animal model
- Human

Good Manufacturing Practice: GMP

- Or “Generate More Paperwork” ...
- Injectable preparations for human (and for animal also) must respect GMP preparation rules
- Guidelines on hardware, software and paperwork validation
- Radiopharmaceuticals are drugs, but up to 4 batches produced every day (vs few batches per year of traditional products)

Physics research in Nuclear Medicine field

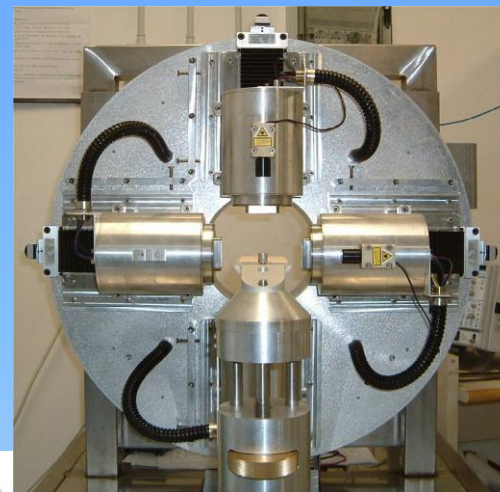
- Nuclide production/optimization by accelerator or generator
- Accelerator construction/maintenance/optimization
- Imaging apparatuses prototypization: PET, SPECT, CT
- Image extraction, based on matter interaction phenomena (theory) and mathematical iterative methods (practice)
- Radiotherapy machines building
- Radiosurveillance, controlled are site planning, personnell control

Preclinical multimodal imaging at IFC

Xalt_{HR}

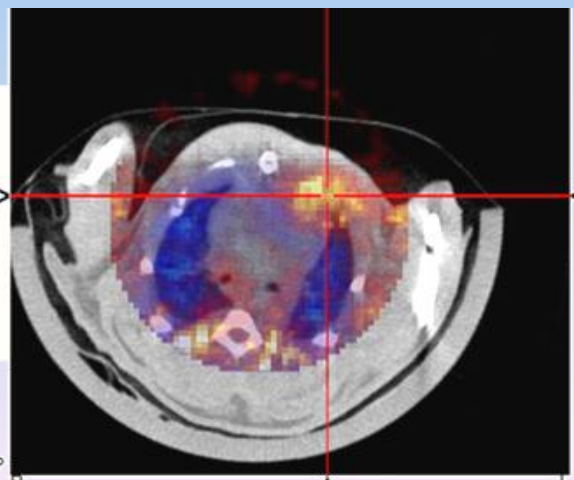


YAP-(S)PET II

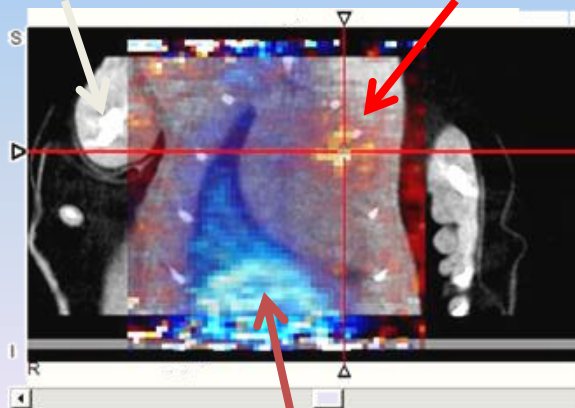


Ingegneria dei Sistemi Elettronici

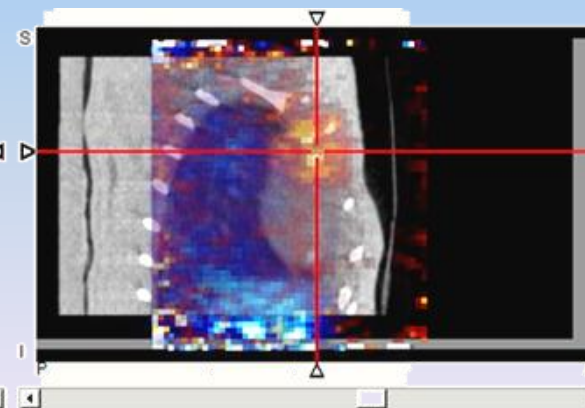
CT



⁶⁸Ga-PET



¹³NH₃-PET



Examples of new prototypes



Mini-cyclotron
(Ron Nutt, ABT)



Automated chemistry



Automated Quality
Control



ABT Molecular Imager, Inc.

Thanks for the attention